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(21) International Application Number: PCT/US00/11085 (22) International Filing Date: 25 April 2000 (25.04.00) (30) Priority Data: 09/300,971 28 April 1999 (28.04.99) US (63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Application US 09/300,971 (CIP) Filed on 28 April 1999 (28.04.99) (71) Applicant (for all designated States except US): THE ADMINISTRATORS OF THE TULANE EDUCATIONAL FUND [US/US]; 1430 Tulane Avenue, New Orleans, LA 70112-2699 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): PHILIPP, Mario, T. [DE/US]; 248 Shaunell Drive, Mandeville, LA 70448 (US). LIANG, Fang, Ting [CN/US]; 7 Lurline Drive, Apt. 9, Covington, LA 70433 (US).	(74) Agents: KODROFF, Cathy, A. et al.; Howson & Howson, Spring House Corporate Center, P.O. Box 457, Spring House, PA 19477 (US). (81) Designated States: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>	
(54) Title: PEPTIDES AND ASSAYS FOR THE DIAGNOSIS OF LYME DISEASE AND COMPOSITIONS USEFUL FOR THE PREVENTION THEREOF		
(57) Abstract A peptide consisting of an invariable 26-amino-acid-long region, named IR ₆ , which is antigenically conserved among strains and species of the <i>B. burgdorferi sensu lato</i> complex, and immunodominant in both human and nonhuman primate hosts is described. This peptide is characterized by the sequence MKKDDQIAAAMVLRGMAKDGGQFALKD [SEQ ID NO: 1]. This peptide is useful for rapid and specific diagnosis of Lyme disease, as are proteins containing this peptide and nucleic acid sequences encoding this peptide and these proteins. Also provided is a novel ELISA, which is characterized by high sensitivity and specificity.		

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PEPTIDES AND ASSAYS FOR THE DIAGNOSIS OF LYME DISEASE AND COMPOSITIONS USEFUL FOR THE PREVENTION THEREOF

This invention was funded in part by the National Institutes of Health Grant Nos. RO1 AI35027 and RR00164. The United States government has certain
5 rights in this invention.

Field of the Invention

The present invention relates generally to the field of diagnostic compositions and methods useful in the diagnosis of Lyme borreliosis.

Background of the Invention

10 The bacterium *Borrelia burgdorferi* (*sensu lato*) is the causative agent of Lyme borreliosis, i.e., Lyme disease. This disease is transmitted by the bite of various species of *Ixodes* ticks carrying the spirochete. The main reservoir of the infection in the United States is the white footed mouse, *Peromyscus leucopus*, and the infection can be transmitted to many mammalian species including dogs, cats, and
15 man [J. G. Donahue, *et al*, Am. J. Trop. Med. Hyg., 36:92-96 (1987); R. T. Green, *et al*, J. Clin. Micro., 26:648-653 (1988)]. Despite the presence of an active immune response, the disease persists for years in patients. Such persistence is postulated to be the result, at least in part, of antigenic variation in the bacterial proteins [J.R. Zhang *et al*, Cell, 89:275-285 (1997)].

20 A number of *Borrelia burgdorferi* proteins and polypeptides have been described in the literature. Such proteins and polypeptides include outer surface proteins A and B (OspA and OspB), flagellin, and other proteins designated P21, P39, P66, and P83 according to their estimated molecular weights [A. G. Barbour *et al*, Infect. Immun., 45:94-100 (1984); W. J. Simpson *et al*, J. Clin. Microbiol., 28:1329-
25 1337 (1990); K. Hansen *et al*, Infect. Immun., 56:2047-2053 (1988); K. Hansen *et al*, Infect. J. Clin. Microbiol., 26:338-346 (1988); B. Wilske *et al*, Zentral. Bakteriол. Parasitenkd. Infektionshkr. Hyg. Abt. 1 Orig. Reihe. A, 263:92-102 (1986); D. W.

Dorward *et al*, J. Clin. Microbiol., 29:1162-1170 (1991); published NTIS US patent application No. 485,551; European patent application No. 465,204, published January 8, 1992; International Patent Application No. PCT/US91/01500, published September 19, 1991; International Patent Application No. PCT/EP90/02282, published July 11, 5 1991; International Patent Application No. PCT/DK89/00248, published May 3, 1990; International patent application No. WO92/00055, published January 9, 1992].

However, the diagnosis of Lyme disease in humans and animals has been compromised by the lack of definitive serology leading to rapid and accurate testing. Current diagnostic tests suffer from low sensitivity and specificity, as 10 illustrated by a recent survey of diagnostic laboratories' performance issued by the Wisconsin State Laboratory of Hygiene [L. Bakken *et al*, J. Clin. Microbiol., 35:537 (1997)].

There is thus a need in the art for a simple, sensitive and specific diagnostic composition and method for early detection of Lyme disease.

15

Summary of the Invention

The present invention satisfies the need in the art by providing methods and compositions which permit rapid and accurate detection of Lyme disease. The methods and compositions of the invention advantageously avoid serologic cross- 20 reactivity with other conditions, including syphilis, chronic arthritis, and multiple sclerosis, from which differential diagnosis was required using prior art methods.

In one aspect, the invention provides a peptide having the amino acid sequence MKKDDQIAAAMVLRGMAKDGGFALKD [SEQ ID NO: 1], termed herein IR₆, which is an invariable region which is immunodominant in mammalian 25 Lyme disease patients.

In another aspect, the invention provides an artificial protein which contains the amino acid sequence of IR₆, or an analog, homolog or fragment thereof. In one embodiment, this protein may be a fusion protein containing IR₆ or a fragment thereof and a fusion partner. In one particularly desirable embodiment, the fusion

protein contains a fragment of IR₆ corresponding to a T cell epitope, wherein the T cell epitope is fused to a one of the other five invariable regions identified herein (IR₁-IR₅).

In another desirable embodiment, the artificial protein contains the
5 amino acid sequence of IR₆ with an N-terminal amino acid suitable for biotinylation.

In yet another aspect, the invention provides a nucleic acid sequence encoding IR₆ peptide or a protein of the invention. In still another aspect, the invention provides a vector comprising a nucleic acid sequence according to the invention under the control of suitable regulatory sequences. In a further aspect, the
10 invention provides a host cell transformed with the vector of the invention.

In a still further aspect, the present invention provides a diagnostic reagent comprising a nucleic acid sequence of the invention and a detectable label which is associated with said sequence.

In yet a further aspect, the invention provides an isolated antibody
15 which is specific for the IR₆ peptide of the invention or a fragment thereof. In still a further aspect, the invention provides a diagnostic reagent comprising the antibody of the invention.

In yet another aspect, the invention provides an anti-idiotypic antibody specific for the anti-IR₆ antibody of the invention and a diagnostic reagent containing
20 the anti-idiotypic antibody with a detectable label linked thereto.

In yet a further aspect, the invention provides a method for diagnosing Lyme disease in a mammal. This method includes the steps of incubating an antigen or antibody of this invention, preferably conventionally labeled for detection, with a sample of biological fluids from a human or an animal to be diagnosed. In the
25 presence of *B. burgdorferi* infection of the human or verterinary patient, an antigen-antibody complex is formed. Subsequently the reaction mixture is analyzed to determine the presence or absence of these antigen-antibody complexes. In a further embodiment, the diagnostic assay employs DNA sequences, preferably anti-sense sequences, of the antigen or fragments thereof, and diagnoses infection by the

presence of sequences in a biological fluid from the patient that hybridizes thereto. Other conventional assay formats may be employed using reagents identified by this invention.

5 In another aspect the invention provides a kit for diagnosing infection with *B. burgdorferi* in a human or a veterinary patient sample which contains at least one antibody capable of binding at least one antigen of this invention of antigenic fragment(s) thereof, or a DNA sequence encoding one or more antigen(s) of this invention or an anti-sense sequence thereof. The antibodies and sequences may be optionally labeled for detection, or a detection system may be included in the kit.

10 In another aspect, the invention provides a therapeutic composition and methods for treating humans and/or animals with Lyme disease. The therapeutic composition contains an antibody, or protein, or fragment as described above and a suitable pharmaceutical carrier.

15 In a further aspect, the invention provides vaccine compositions and methods of vaccinating a human or animal patient against Lyme Disease by use of these above-described compositions. The vaccine composition may contain the IR₆ protein, fragments thereof, fusion proteins or mixtures of proteins as described above with a pharmaceutically acceptable carrier. More preferably, the vaccine compositions contain fusion proteins composed of IR₆, or a fragment thereof corresponding to a T
20 cell epitope, fused to a partner such as IR₁ - IR₅.

In yet a further aspect, the invention provides vaccine compositions and methods of vaccinating a human or animal patient against Lyme Disease by use of nucleic acid compositions, e.g., DNA vaccines. The compositions contain an effective amount of a DNA sequence encoding at least one IR_{1,5} or IR₆ peptide or protein of
25 this invention and a pharmaceutically acceptable carrier.

Other aspects and advantages of the present invention are described further in the following detailed description of the preferred embodiments thereof.

Brief Description of the Drawings

Fig. 1 illustrates an antibody response to P7-1 in monkeys. Serum samples from two rhesus macaques that were infected with the JD1 strain of *B. burgdorferi* (J831 and L131) and two that were infected with B31 strain spirochetes (L457 and M021) were serially collected. The antibody (Ab) level was measured by ELISA using P7-1 as antigen (Ag). The baseline represents the mean ELISA optical density (OD) of serum samples collected from all of the animals before infection plus 3 standard deviations (SDs).

Fig. 2 provides a comparison among deduced amino acid sequences of P7-1 and cassette segments from VlsE of strains B31 and 297 of *B. burgdorferi*. The B31 cassette segment is confined between the repeat sequence EGAIKG [SEQ ID NO: 2], singly underlined. Variable regions (VR₁₋₃) are doubly underlined and invariable regions (IR₁₋₆) are shaded. Sequence alignment was obtained with the pam250 algorithm (Eastman Kodak Co., New Haven, CT). Identical amino acids are indicated with asterisks and sequence gaps with dashes; nonidentical residues are shown as letters following the single letter code.

Figs. 3A and 3B are bar graphs which illustrate the antigenicity of IR₆ in monkeys (A) and mice (B). Serum samples were collected from monkeys or mice at 0 (pre) and 4-6 wk post infection (PI; post). Animals were infected with the JD1 strain of *B. burgdorferi* (monkeys J200, J415, J748, J831, K205 and L131), the B31 strain (monkeys L457, L549, M021 and M581; mice 184, 191, 194 and 196), or the Sh-2-82 strain (mice 219, 220, 224, 288, 289 and 290). Ab levels were assessed by the C₆ ELISA.

Figs. 4A and 4B are graphs which illustrate the immunodominance of IR₆ in monkeys and humans. Serum samples were obtained from (A) monkeys inoculated 4-6 wk PI with spirochetes of the JD1 strain of *B. burgdorferi* (J831 and L131) or the B31 strain (L457 and M021), and from (B) humans with an acute *B. burgdorferi* infection (91-1222 and 91-1348) or a chronic infection (91-0532 and 91-0533). Ab levels to P7-1 were assessed in the presence of increasing concentrations of the C₆ peptide by the competitive ELISA procedure.

Detailed Description of the Invention

The present invention satisfies the need in the art by providing methods and compositions which permit rapid and accurate detection of Lyme disease in humans. The methods and compositions of the invention advantageously avoid
5 serologic cross-reactivity with other conditions, including syphilis, chronic arthritis, and multiple sclerosis, from which differential diagnosis was required using prior art methods. The invention is also useful in detection of Lyme disease in non-human mammals, including dogs, horses, and cows, among others.

In a currently preferred embodiment, the invention provides a peptide-
10 based ELISA, which in terms of simplicity, specificity and sensitivity, is superior to current serologic diagnostic methods for Lyme disease. Further, the ELISA of the invention is also useful in serum samples that contain anti-OspA antibodies, permitting diagnosis of Lyme disease in mammals who have been exposed (either naturally or through vaccination) with *B. burgdorferi*, the causative agent of Lyme disease.

The methods and compositions of the invention utilize a peptide having
15 the amino acid sequence MKKDDQIAAAMVLRGMAKDGQFALKD [SEQ ID NO: 1], which provides the specificity for Lyme disease which is an advantage of the invention. Also provided by the present invention are proteins containing this peptide, as well as antibodies directed thereto and nucleic acid sequences encoding
20 these peptides and/or proteins for use in diagnostic, therapeutic and prophylactic compositions and methods for the treatment or prevention of Lyme Disease. The invention provides advantages over the use of other *Borrelia* proteins and antibodies in known compositions and methods for this purpose.

I. The Invariable Region Peptides, Proteins and Nucleic Acids of the Invention

25 The present invention relates to six antigenic peptides which correspond to invariable regions (IR₁₋₆), found within the variable domain of the variable surface antigen of *B. burgdorferi* (VlsE). These IRs are conserved among strains and genospecies of the *B. burgdorferi* sensu lato complex. Surprisingly, unlike the variable regions of several other major proteins, which are not antigenic in natural

infections, the most conserved of the IRs, IR₆, is immunodominant in Lyme disease patients and in animals infected with *B. burgdorferi*. IR₆ is exposed on the surface of VlsE, as assessed by immunoprecipitation experiments, but is inaccessible to antibody on the spirochete's outer membrane, as demonstrated by immunofluorescence and *in vitro* killing assays.

Thus, the present invention provides six novel peptides and fragments thereof which may be used in a variety of diagnostic assays and compositions. Of these peptides, the IR₆ peptide and proteins containing this peptide or fragments thereof are particularly well suited for use in specific diagnosis of Lyme disease. The peptides IR_{1,5} may also be used for such a purpose. The peptides IR_{1,5} described herein and proteins containing one or more of these peptides or fragments thereof are particularly well suited for use in therapeutic and pharmaceutical compositions for the treatment of Lyme disease. IR₆ may also be used for such purposes.

A. Protein and Peptide Sequences

For convenience throughout this specification, reference will be made to "IR_{1,5} or IR₆ peptides and proteins", but it will be understood that this term encompasses the fragments, analogs, modified peptides and proteins, fusion proteins, and other amino acid constructs of the invention, except where otherwise specified.

In one aspect, the invention provides novel peptides having the amino sequences provided below, utilizing standard single letter amino acid codes.

IR₁: GNAAIGDVV [SEQ ID NO: 3]
 IR₂: SVNGIAKGIGIVDAA [SEQ ID NO: 4]
 IR₃: AGKLFVK [SEQ ID NO: 5]
 IR₄: DAGKAAA VAAVSGEQILKAIVHAA [SEQ ID
 NO: 6],
 IR₅: ATNPIDAAIG [SEQ ID NO: 7] and
 IR₆: MKKDDQIAAAMVLRGMAKDGGQFALKD [SEQ ID
 NO: 1].

These peptide antigens may be isolated in a form substantially free from other proteinaceous and non-proteinaceous materials of the microorganism

and the tick vector. The antigens may be isolated from the spirochete and further purified using any of a variety of conventional methods including: liquid chromatography such as normal or reversed phase, using HPLC, FPLC and the like; affinity chromatography (such as with inorganic ligands or monoclonal antibodies);
5 size exclusion chromatography; immobilized metal chelate chromatography; gel electrophoresis; and the like. One of skill in the art may select the most appropriate isolation and purification techniques without departing from the scope of this invention.

Alternatively, the peptides and proteins of the invention,
10 described below, may be produced recombinantly following conventional genetic engineering techniques [see e.g., Sambrook et al, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratories, Cold Spring Harbor, NY and the detailed description of making the proteins below]. In still another alternative, the peptides and proteins of the invention may be produced using conventional chemical synthesis
15 techniques, such as those described in G. Barony and R.B. Merrifield, The Peptides: Analysis, Synthesis & Biology, Academic Press, pp. 3-285 (1980), among others. The term "artificial" is used herein to denote the preparation of the construct (e.g., a peptide, protein, nucleic acid, or antibody of the invention) by chemical synthesis, recombinant technology, or other similar means.

20 The present invention further provides analogs, fragments, and mutant peptides, as well as proteins containing one or more of IR₁₋₆, or such analogs, fragments or mutants, as described below.

i. Analogs and Modified Peptide and Protein Antigens

Analogues or modified versions of the peptides IR₁₋₆ are
25 provided. Typically, analogs differ from the specifically identified proteins by only one to four codon changes. Examples include polypeptides with minor amino acid variations from the illustrated partial amino acid sequence of, for example, IR₂ having conservative amino acid replacements. Conservative replacements are those that take place within a family of amino acids that are related in their side chains and chemical
30 properties. Also provided are homologs of the proteins of the invention which are

characterized by having at least 90% identity, and more preferably 95-99% identity with IR₁ the sequences of the vls-like proteins. Based on the sequence information provided herein, one of skill in the art can readily obtain full-length homologs and analogs.

5 As known in the art, "homology" or "identity" means the degree of sequence relatedness between two peptide or two nucleotide sequences as determined by the identity of the match between two lengths of such sequences. Both identity and homology can be readily calculated by methods extant in the prior art [See, e.g., COMPUTATIONAL MOLECULAR BIOLOGY, Lesk, A.M., ed., Oxford University Press, New York, (1988); BIOCOMPUTING: INFORMATICS AND GENOME PROJECTS, Smith, D.W., ed., Academic Press, New York, (1993); COMPUTER ANALYSIS OF SEQUENCE DATA, PART I, Griffin, A.M., and Griffin, H.G., eds., Humana Press, New Jersey, (1994); SEQUENCE ANALYSIS IN MOLECULAR BIOLOGY, von Heinje, G., Academic Press, (1987); and SEQUENCE ANALYSIS PRIMER, Gribskov, M. and Devereux, J., eds., M Stockton Press, New York, (1991)]. While there exist a number of methods to measure identity and homology between two nucleotide sequences, the terms "identity", "similarity" and homology are well known to skilled artisans [H. Carillo and D. Lipton, SIAM J. Applied Math., 48:1073 (1988)]. Methods commonly employed to determine identity or homology between two sequences include, but are not limited to, those disclosed in Guide to Huge Computers, Martin J. Bishop, ed., Academic Press, San Diego, 1994, and H. Carillo and D. Lipton, SIAM J. Applied Math., 48:1073 (1988). Preferred methods to determine identity or homology are designed to give the largest match between the two sequences tested. Methods to determine identity and similarity are codified in computer programs. Preferred computer program methods to determine identity and homology between two sequences include, but are not limited to, the algorithm BESTFIT from the GCG program package [J. Devereux et al., Nucl. Acids Res., 12(1):387 (1984)], the related MACVECTOR program (Oxford), and the FASTA (Pearson) programs, which may

be used at default settings or modified settings such as determined to be suitable by one of skill in the art.

An IR₁₋₅ or IR₆ peptide or protein of the present invention may also be modified to increase its immunogenicity. For example, the antigen may be coupled to a chemical compounds or immunogenic carriers, provided that the coupling does not interfere with the desired biological activity of either the antigen or the carrier. For a review of some general considerations in coupling strategies, see Antibodies, A Laboratory Manual, Cold Spring Harbor Laboratory, ed. E. Harlow and D. Lane (1988). Useful immunogenic carriers known in the art, include, without limitation, keyhole limpet hemocyanin (KLH); bovine serum albumin (BSA), ovalbumin, PPD (purified protein derivative of tuberculin); red blood cells; tetanus toxoid; cholera toxoid; agarose beads; activated carbon; or bentonite. Useful chemical compounds for coupling include, without limitation, dinitrophenol groups and arsonilic acid.

The IR₁₋₆ peptides and proteins of the invention may also be modified by other techniques, such as by denaturation with heat and/or SDS. Alternatively, the peptides and proteins of the invention may be modified to provide an additional N- or C-terminal amino acid sequence suitable for biotinylation, e.g., cysteine or lysine.

ii. *Fragments/Deletion Mutants*

Further encompassed by this invention are additional fragments of the IR₁₋₅ peptides, the IR₆ peptide or the other proteins identified herein. Such fragments are desirably characterized by having a biological activity similar to that displayed by the complete protein, including, e.g., the ability to induce antibodies to the causative agent of Lyme Disease. These fragments may be designed or obtained in any desired length, including as small as about 5 to 8 amino acids in length. Such a fragment may represent an epitope of the protein.

For example, one particularly desirable fragment of the invention is a T cell epitope located within a peptide of the invention, e.g., IR₆. Such a T cell epitope may be readily identified using available computer modelling programs.

5 Optionally, the peptides of the invention may be modified to create deletion mutants, for example, by truncation at the amino or carboxy termini, or by elimination of one or more amino acids. Still other modified fragments of IR₁₋₅ or IR₆ may be prepared by any number of now conventional techniques to improve production thereof, to enhance protein stability or other
10 characteristics, e.g. binding activity or bioavailability, or to confer some other desired property upon the protein. Other useful fragments of these polypeptides may be readily prepared by one of skill in the art using known techniques, such as deletion mutagenesis and expression.

iii. *Fusion or Multimeric Proteins and Compositions*

15 The IR₁₋₅ or IR₆ peptides of the present invention, or fragments of it, may also be constructed, using conventional genetic engineering techniques as part of a larger and/or multimeric protein or protein compositions. In one currently preferred embodiment, a fusion protein is composed of an IR₆ fragment corresponding to a T cell epitope, which is fused to a peptide selected from among the
20 IR₁₋₅ peptides of the invention.

 The IR₁₋₅ and IR₆ peptides and proteins of this invention may be used in combination with *B. burgdorferi* outer surface proteins, such as OspA and OspB, or various fragments of these may be used in combination with each other. In such combination, the antigen may be in the form of a fusion protein. Thus, an
25 antigen of the invention (e.g., IR₆ or a fragment thereof) may be optionally fused to a selected polypeptide or protein, e.g. *Borrelia* antigens OspA and OspB, other *Borrelia* antigens, and/or proteins or polypeptides derived from other microorganisms. For example, a peptide or polypeptide of this invention may be fused at its N-terminus or C-terminus to OspA polypeptide, or OspB polypeptide or to a
30 non-OspA non-OspB polypeptide or combinations thereof. OspA and OspB

polypeptides which may be useful for this purpose include polypeptides identified by the prior art [see, e.g. PCT/US91/04056] and variants thereof. Non-OspA, non-OspB polypeptides which may be useful for this purpose include polypeptides of the invention and those identified by the prior art, including, the *B. burgdorferi*, flagella-associated protein and fragments thereof, other *B. burgdorferi* proteins and fragments thereof, and non-*B. burgdorferi* proteins and fragments thereof.

These fusion proteins are constructed for use in the methods and compositions of this invention. These fusion proteins or multimeric proteins may be produced recombinantly, or may be synthesized chemically. They also may include the peptides and proteins of this invention fused or coupled to moieties other than amino acids, including lipids and carbohydrates. Further, antigens of this invention may be employed in combination with other *Borrelia* vaccinal agents described by the prior art, as well as with other species of vaccinal agents derived from other viruses. Such proteins are effective in the prevention, treatment and diagnosis of Lyme disease as caused by a wide spectrum of *B. burgdorferi* isolates.

A protein composition which may be a preferred alternative to the fusion proteins described above is a cocktail (i.e., a simple mixture) containing an IR₆ peptide or protein, or different mixtures of the IR₁₋₅ and IR₆ peptides and proteins of this invention.

In still another aspect, the peptide and proteins of the invention may be provided with a detectable label, such as are described in detail below.

iv. Salts

A peptide or protein antigen of the present invention may also be used in the form of a pharmaceutically acceptable salt. Suitable acids and bases which are capable of forming salts with the polypeptides of the present invention are well known to those of skill in the art, and include inorganic and organic acids and bases.

B. Nucleic Acid Sequences

The present invention provides mammalian nucleic acid sequences encoding the IR₁₋₅ and IR₆ peptides and proteins of the invention, as defined above. Also provided are nucleic acid sequences complementary to the coding strand, e.g., the anti-sense strand, and corresponding RNA sequences.

Allelic variants of these sequences within a species (i.e., sequences containing some individual nucleotide differences from a more commonly occurring sequence within a species, but which nevertheless encode the same protein or a protein with the same function) may also be readily obtained given the knowledge of the nucleic acid sequence provided by this invention (see Fig. 2).

The present invention further encompasses nucleic acid sequences capable of hybridizing under stringent conditions [see, J. Sambrook et al, Molecular Cloning: A Laboratory Manual, 2d ed., Cold Spring Harbor Laboratory (1989)] to the sequences of the invention, their anti-sense strands, or biologically active fragments thereof. An example of a highly stringent hybridization condition is hybridization at 2XSSC at 65°C, followed by a washing in 0.1XSSC at 65°C for an hour. Alternatively, an exemplary highly stringent hybridization condition is in 50% formamide, 4XSSC at 42°C. Moderately high stringency conditions may also prove useful, e.g., hybridization in 4XSSC at 55°C, followed by washing in 0.1XSSC at 37°C for an hour. An alternative exemplary moderately high stringency hybridization condition is in 50% formamide, 4XSSC at 30°C.

According to the invention, the nucleic acid sequences may be modified. Utilizing the sequence data provided herein, it is within the skill of the art to obtain or prepare synthetically or recombinantly other polynucleotide sequences, or modified polynucleotide sequences, encoding the full-length proteins or useful fragments of the invention. Such modifications at the nucleic acid level include, for example, modifications to the nucleotide sequences which are silent or which change the amino acids, e.g. to improve expression or secretion. Also included are allelic variations, caused by the natural degeneracy of the genetic code.

Also encompassed by the present invention are mutants of the IR₁₋₅ and IR₆ peptides and proteins provided herein. Such mutants include amino terminal, carboxy terminal or internal deletions, which substantially retain the antigenicity of the full-length IR₁₋₅ and IR₆ or other proteins or fragments. Such a truncated, or deletion mutant may be expressed for the purpose of affecting the activity of the full-length or wild-type gene or gene fragments.

Thus, the invention provides fragments that encode a desirable fragment of IR₁₋₅ or IR₆, e.g., a T cell epitope. Generally, these oligonucleotide fragments are at least 15 nucleotides in length. However, oligonucleotide fragments of varying sizes may be selected as desired. Such fragments may be used for such purposes as performing polymerase chain reaction (PCR), e.g., on a biopsied tissue sample.

The nucleic acid sequences of the invention may be obtained, in whole or in part, from natural sources and isolated from the cellular materials with which they are naturally associated. For example, the sequences encoding IR₁₋₅ and IR₆ were originally isolated from a fragment, designated 7-1, from *Borrelia garinii* strain IP90, inserted in pBluescript II plasmid, was transformed in *E. coli* and deposited with the American Type Culture Collection, Manassas, Virginia ("ATCC") on June 27, 1997 under Accession No. 98478. 7-1 is described in detail in International Patent Publication No. WO99/00413, published January 7, 1999, and incorporated herein by reference. However, the sequences of the invention may be isolated from other suitable sources by conventional uses of polymerase chain reaction or cloning techniques such as those described in conventional texts such as Sambrook et al, cited above.

More desirably, these nucleic acid sequences of the invention may be constructed recombinantly using conventional genetic engineering or chemical synthesis techniques or PCR, and the like by utilizing the information provided herein. These nucleic acid sequences are useful for a variety of diagnostic, prophylactic and therapeutic uses. Advantageously, the nucleic acid sequences are useful in the development of diagnostic probes and antisense probes for use in the detection and

diagnosis of Lyme disease by utilizing a variety of known nucleic acid assays, e.g., Northern and Southern blots, polymerase chain reaction (PCR), and other assay techniques known to one of skill in the art. When used in diagnostic applications, the nucleic acid sequences of the invention may optionally be associated with a detectable label, such as are described in detail below. The nucleic acid sequences of this invention are also useful in the production of the peptides and proteins of the invention.

II. Methods of Making Antigens and Nucleic Acid Sequences of the Invention

A. *Chemical Synthesis*

The peptides, proteins, and nucleic acid sequences of the invention may be prepared conventionally by resort to known chemical synthesis techniques, e.g., solid-phase chemical synthesis, such as described by Merrifield, J. Amer. Chem. Soc., 85:2149-2154 (1963), and J. Stuart and J. Young, Solid Phase Peptide Synthesis, Pierce Chemical Company, Rockford, IL (1984), or detailed in the examples below. A variety of methods for producing the above-identified modifications of the peptides or nucleic acid sequences are known and may be selected by one of skill in the art.

B. *Expression In Vitro*

To produce the recombinant IR₁₋₅ and IR₆ peptides and proteins of the invention, the DNA sequences of the invention are inserted into a suitable expression system. Desirably, a recombinant molecule or vector is constructed in which the polynucleotide sequence encoding the selected protein, e.g., an IR₆ peptide or protein, is operably linked to a heterologous expression control sequence permitting expression of the protein. Numerous types of appropriate expression vectors are known in the art for protein expression, by standard molecular biology techniques. Such vectors are selected from among conventional vector types including insects, e.g., baculovirus expression, or yeast, fungal, bacterial or viral expression systems. Other appropriate expression vectors, of which numerous types are known in the art, can also be used for this purpose. Methods for obtaining such

expression vectors are well-known. See, Sambrook et al, Molecular Cloning. A Laboratory Manual, 2d edition, Cold Spring Harbor Laboratory, New York (1989); Miller et al, Genetic Engineering, 8:277-298 (Plenum Press 1986) and references cited therein.

5 Suitable host cells or cell lines for transfection by this method include bacterial cells. For example, the various strains of *E. coli* (e.g., HB101, MC1061, and strains used in the following examples) are well-known as host cells in the field of biotechnology. Various strains of *B. subtilis*, *Pseudomonas*, *Streptomyces*, and other bacilli and the like are also be employed in this method.

10 Mammalian cells, such as human 293 cells, Chinese hamster ovary cells (CHO), the monkey COS-1 cell line or murine 3T3 cells derived from Swiss, Balb-c or NIH mice are used. Another suitable mammalian cell line is the CV-1 cell line. Still other suitable mammalian host cells, as well as methods for transfection, culture, amplification, screening, production, and purification are known
15 in the art. [See, e.g., Gething and Sambrook, Nature, 293:620-625 (1981), or alternatively, Kaufman et al, Mol. Cell. Biol., 5(7):1750-1759 (1985) or Howley et al, U. S. Patent 4,419,446].

 Many strains of yeast cells known to those skilled in the art are also available as host cells for expression of the polypeptides of the present invention.
20 Other fungal cells may also be employed as expression systems. Alternatively, insect cells such as *Spodoptera frugiperda* (Sf9) cells may be used.

 Thus, the present invention provides a method for producing a recombinant IR₁₋₅ or IR₆ peptide or protein, which involves transfecting, e.g., by conventional means such as electroporation, a host cell with at least one expression
25 vector containing a polynucleotide of the invention under the control of a transcriptional regulatory sequence. The transfected or transformed host cell is then cultured under conditions that allow expression of the protein. The expressed protein is recovered, isolated, and optionally purified from the cell (or from the culture medium, if expressed extracellularly) by appropriate means known to one of skill in
30 the art.

For example, the proteins are isolated in soluble form following cell lysis, or extracted using known techniques, e.g., in guanidine chloride. If desired, the proteins or fragments of the invention are produced as a fusion protein. Such fusion proteins are those described above. Alternatively, for example, it may be desirable to produce fusion proteins to enhance expression of the protein in a selected host cell, to improve purification, or for use in monitoring the presence of the desired peptide or protein, e.g., IR₆, in tissues, cells or cell extracts. Suitable fusion partners for the peptides and proteins of the invention are well known to those of skill in the art and include, among others, β -galactosidase, glutathione-S-transferase, and poly-histidine.

C. *Expression In Vivo*

Alternatively, where it is desired that the IR_{1,5} or IR₆ peptide or protein of the invention be expressed *in vivo*, e.g., to induce antibodies, or as a DNA vaccine, an appropriate vector for delivery is readily selected by one of skill in the art. Exemplary vectors for *in vivo* gene delivery are readily available from a variety of academic and commercial sources, and include, e.g., adeno-associated virus [see, e.g., International patent application Nos. WO91/18088 and WO95/34670], adenovirus vectors [M. Kay et al, Proc. Natl. Acad. Sci. USA, 91:2353 (1994); S. Ishibashi et al, J. Clin. Invest., 92:883 (1993)], or other viral vectors, e.g., various poxviruses, vaccinia, etc. Methods for insertion of a desired coding sequence, e.g., the sequence encoding the IR₆ peptide, and obtaining *in vivo* expression of the encoded protein, are well known to those of skill in the art.

III. Antibodies of the Invention

The present invention also provides antibodies capable of recognizing and binding the IR_{1,5} and IR₆ peptides of this invention, including antibodies derived from mixtures of such antigens or fragments thereof. These antibodies are useful in diagnosis of Lyme disease and in therapeutic compositions for treating humans and/or animals that test positive for, or, prior to testing, exhibit symptoms of, Lyme Disease. The antibodies are useful in diagnosis alone (e.g., those antibodies raised using IR₆

peptides and proteins) or in combination with antibodies to other antigens of this invention as well as antibodies to other known *B. burgdorferi* antigens. These antibodies, particularly those generated using IR₁₋₅, are also useful in passive vaccine compositions.

5 The antibodies of this invention are generated by conventional means utilizing the isolated, recombinant or modified antigens of this invention, or mixtures of such antigens or antigenic fragments. For example, polyclonal antibodies are generated by conventionally stimulating the immune system of a selected animal or human with the isolated antigen or mixture of antigenic proteins or peptides of this invention, allowing the immune system to produce natural antibodies thereto, and
10 collecting these antibodies from the animal or human's blood or other biological fluid.

 Monoclonal antibodies (MAbs) directed against an IR₁₋₅ or IR₆ peptide or protein of the invention may also be generated. Hybridoma cell lines expressing desirable MAbs are generated by well-known conventional techniques, e.g. Kohler
15 and Milstein and the many known modifications thereof. Similarly desirable high titer antibodies are generated by applying known recombinant techniques to the monoclonal or polyclonal antibodies developed to these antigens [see, e.g., PCT Patent Application No. PCT/GB85/00392; British Patent Application Publication No. GB2188638A; Amit et al., Science, 233:747-753 (1986); Queen et al., Proc. Nat'l.
20 Acad. Sci. USA, 86:10029-10033 (1989); PCT Patent Application No. PCT/WO9007861; and Riechmann et al., Nature, 332:323-327 (1988); Huse et al., Science, 246:1275-1281 (1988)].

 Given the disclosure contained herein, one of skill in the art may generate chimeric, humanized or fully human antibodies directed against an IR₁₋₅ or
25 IR₆ peptide or protein of the invention by resort to known techniques by manipulating the complementarity determining regions of animal or human antibodies to the antigen of this invention. See, e.g., E. Mark and Padlin, "Humanization of Monoclonal Antibodies", Chapter 4, The Handbook of Experimental Pharmacology, Vol. 113, The Pharmacology of Monoclonal Antibodies, Springer-Verlag (June, 1994).

Alternatively, the antigens may be assembled as multi-antigenic complexes [see, e.g., European Patent Application 0339695, published November 2, 1989] or as simple mixtures of antigenic proteins/peptides and employed to elicit high titer antibodies capable of binding the selected antigen(s) as it appears in the biological fluids of an infected animal or human.

Further provided by the present invention are anti-idiotypic antibodies (Ab2) and anti-anti-idiotypic antibodies (Ab3). Ab2 are specific for the target to which anti-IR₁₋₅ or anti-IR₆ antibodies (Ab1) of the invention bind and Ab3 are similar to Ab1 in their binding specificities and biological activities [see, e.g., M. Wettendorff et al., "Modulation of anti-tumor immunity by anti-idiotypic antibodies." In Idiotypic Network and Diseases, ed. by J. Cerny and J. Hiernaux J, Am. Soc. Microbiol., Washington DC: pp. 203-229, (1990)]. These anti-idiotypic and anti-anti-idiotypic antibodies are produced using techniques well known to those of skill in the art. Such anti-idiotypic antibodies (Ab2) can bear the internal image of the antigens, and are thus useful for the same purposes as the IR₁₋₅ and IR₆ peptides and proteins of the invention.

In general, polyclonal antisera, monoclonal antibodies and other antibodies which bind to the selected antigen (Ab1) are useful to identify epitopes of IR₁₋₅ and IR₆ and to separate these peptides and proteins from contaminants in tissue (e.g., in chromatographic columns and the like), and in general as research tools and as starting materials essential for the development of other types of antibodies described above. Anti-idiotypic antibodies (Ab2) are useful for binding the same target and thus may be used in place of the original antigen.

For use in diagnostic assays, the antibodies are associated with conventional labels which are capable, alone or in concert with other compositions or compounds, of providing a detectable signal. Where more than one antibody is employed in a diagnostic method, the labels are desirably interactive to produce a detectable signal. Most desirably, the label is detectable visually, e.g. colorimetrically. A variety of enzyme systems have been described in the art which will operate to reveal a colorimetric signal in an assay. As one example, glucose oxidase (which uses

glucose as a substrate) releases peroxide as a product. Peroxidase, which reacts with peroxide and a hydrogen donor such as tetramethyl benzidine (TMB), produces an oxidized TMB that is seen as a blue color. Other examples include horseradish peroxidase (HRP) or alkaline phosphatase (AP), and hexokinase in conjunction with glucose-6-phosphate dehydrogenase which reacts with ATP, glucose, and NAD⁺ to yield, among other products, NADH that is detected as increased absorbance at 340 nm wavelength. Other label systems that may be utilized in the methods of this invention are detectable by other means, e.g., colored latex microparticles [Bangs Laboratories, Indiana] in which a dye is embedded may be used in place of enzymes to form conjugates with the antibodies and provide a visual signal indicative of the presence of the resulting complex in applicable assays. Still other labels include fluorescent compounds, radioactive compounds or elements. Detectable labels for attachment to antibodies useful in diagnostic assays of this invention may be easily selected from among numerous compositions known and readily available to one skilled in the art of diagnostic assays. The methods and antibodies of this invention are not limited by the particular detectable label or label system employed. Suitably, these detectable systems may also be utilized in connection with diagnostic reagents composed of the peptides, proteins, and nucleic acid sequences of the invention.

IV. Diagnostic Methods and Assays

The present invention provides reliable and accurate methods of diagnosing Lyme disease in mammals. While particularly desirable for use in humans and dogs, other mammalian animals may be diagnosed using the methods described herein. These diagnostic methods are useful for diagnosing humans or other mammals exhibiting the clinical symptoms of, or suspected of having, Lyme disease. The IR₆ peptides, proteins and nucleic acids of the invention are particularly well suited for use in the diagnostic methods and compositions of the invention. For convenience, reference will be made to IR₆ throughout this and the following section. However, it will be understood that the IR₁₋₅ peptides, proteins and nucleic acids may be useful in these methods.

In one embodiment, this diagnostic method involves detecting the presence of naturally occurring *B. burgdorferi* antibodies which are produced by the infected human or veterinary patient's immune system in its biological fluids, and which are capable of binding to the antigens of this invention or combinations thereof.

5 This method comprises the steps of incubating an IR₆ peptide or protein of this invention with a sample of biological fluids from the patient. Such a sample may be blood, a blood product (e.g., plasma or serum), or another fluid, e.g., tears or urine. A suitable sample may be readily selected based upon the assay format desired. Antibodies present in the fluids as a result of *B. burgdorferi* infection will form an
10 antibody-antigen complex with the antigen. Subsequently the reaction mixture is analyzed to determine the presence or absence of these antigen-antibody complexes. The step of analyzing the reaction mixture comprises contacting the reaction mixture with a labeled specific binding partner for the antibody.

In one embodiment of the method, the IR₆ peptide or protein, or a
15 mixture of the peptides and proteins of the invention is electro- or dot-blotted onto nitrocellulose paper. Subsequently, the biological fluid (e.g. serum or plasma) is incubated with the blotted antigen, and antibody in the biological fluid is allowed to bind to the antigen(s). The bound antibody is then detected by standard immunoenzymatic methods.

20 In another embodiment of the method, latex beads are conjugated to the antigen(s) of this invention. Subsequently, the biological fluid is incubated with the bead/protein conjugate, thereby forming a reaction mixture. The reaction mixture is then analyzed to determine the presence of the antibodies.

In another embodiment, the diagnostic method of the invention involves
25 detecting the presence of the naturally occurring IR₆ antigen(s) itself in its association with the *B. burgdorferi* pathogen in the biological fluids of an animal or human infected by the pathogen. This method includes the steps of incubating an antibody of this invention (e.g. produced by administering to a suitable human and/or animal an antigen of this invention), preferably conventionally labelled for detection, with a
30 sample of biological fluids from a human or an animal to be diagnosed. In the

presence of *B. burgdorferi* infection of the human or animal patient, an antigen-antibody complex is formed (specific binding occurs). Subsequently, excess labeled antibody is optionally removed, and the reaction mixture is analyzed to determine the presence or absence of the antigen-antibody complex and the amount of label
5 associated therewith.

Assays employing a protein antigen of the invention can be heterogenous (i.e., requiring a separation step) or homogenous. If the assay is heterogenous, a variety of separation means can be employed, including centrifugation, filtration, chromatography, or magnetism.

10 One preferred assay for the screening of a patient sample is an enzyme linked immunosorbant assay, i.e., an ELISA. For use in an ELISA, the sample is preferably plasma or serum, although another biological fluid may be used. Typically in an ELISA, the isolated antigen(s) of the invention is adsorbed to the surface of a microtiter well directly or through a capture matrix (i.e., antibody). Residual protein-
15 binding sites on the surface are then blocked with an appropriate agent, such as bovine serum albumin (BSA), heat-inactivated normal goat serum (NGS), or BLOTTO (a buffered solution of nonfat dry milk which also contains a preservative, salts, and an antifoaming agent). The well is then incubated with a biological sample suspected of containing specific anti-*B. burgdorferi* antibody. The sample can be
20 applied neat, or more often, it can be diluted, usually in a buffered solution which contains a small amount (0.1-5.0% by weight) of protein, such as BSA, NGS, or BLOTTO. After incubating for a sufficient length of time to allow specific binding to occur, the well is washed to remove unbound protein and then incubated with labeled anti-human immunoglobulin (α HuIg) or labeled antibodies to other species, e.g.,
25 dogs. The label can be chosen from a variety of enzymes, including horseradish peroxidase (HRP), β -galactosidase, alkaline phosphatase, and glucose oxidase, as described above. Sufficient time is allowed for specific binding to occur again, then the well is washed again to remove unbound conjugate, and the substrate for the enzyme is added. Color is allowed to develop and the optical density of the contents
30 of the well is determined visually or instrumentally.

In one currently preferred embodiment, the invention provides a novel peptide-based ELISA for the diagnosis of Lyme Disease which is a simple, sensitive, and precise assay which can be performed in OspA-vaccinated subjects. Early data showed this assay to have an overall sensitivity of 90%, with a specificity of about 99%, a positive predictive value of about 99% and a negative predictive value of about 89%. Thus, the ELISA of the invention is performed in a ninety-six-well ELISA plate or equivalent solid phase coated with streptavidin or an equivalent biotin-binding compound at an optimal concentration in an alkaline coating buffer and incubated at 4°C overnight. After two 3 minute washes with standard washing buffers such as 10 mM sodium phosphate, 150 mM NaCl and 0.1% Tween-20, pH 7.4 (PBST), an optimal concentration of a biotinylated form of a composition/antigen of this invention dissolved in a conventional blocking buffer such as PBST supplemented with 5% nonfat dry milk, is applied to each well. In a preferred embodiment, the ELISA utilizes the IR6 peptide of the invention or a compound containing this peptide, e.g., the C₆ peptide. After 2 hours of incubation with shaking and three washes as above, a sample of biological fluids at the appropriate dilution, obtained from a human or an animal to be diagnosed, is added to each well. The plate is incubated with shaking for 1 hour at room temperature (RT) and then washed as before. Each well then receives an optimal concentration of an appropriate anti-immunoglobulin antibody that is conjugated to an enzyme or other label by standard procedures and is dissolved in blocking buffer. After one additional hour of incubation followed by four washes with PBST for 3, 4, 5 and 6 minutes, respectively, a solution composed of an appropriate chromogen or indicator is added and color is allowed to develop for 10 minutes. The enzyme reaction is appropriately stopped and optical density is measured at the appropriate wave length. The cutoff OD value may be defined as the mean OD + 3 standard deviations (SDs) of at least 50 serum samples collected from individuals from an area where Lyme disease is not endemic, or by other such conventional definitions. Additional details of the ELISA are provided in the examples below. See, particularly, Example 1G.

Further, MAbs or other antibodies of this invention which are capable of binding to the antigen(s) can be bound to ELISA plates. In another diagnostic method, the biological fluid is incubated on the antibody-bound plate and washed. Detection of any antigen-antibody complex, and qualitative measurement of the labeled MAb is performed conventionally, as described above.

Other useful assay formats include the filter cup and dipstick. In the former assay, an antibody of this invention is fixed to a sintered glass filter to the opening of a small cap. The biological fluid or sample (5 mL) is worked through the filter. If the antigen is present (i.e., *B. burgdorferi* infection), it will bind to the filter which is then visualized through a second antibody/detector. The dipstick assay involves fixing an antigen or antibody to a filter, which is then dipped in the biological fluid, dried and screened with a detector molecule.

Other diagnostic assays can employ the antigen(s) or fragments of this invention as nucleic acid probes or as anti-sense sequences, which can identify the presence of infection in the biological fluid by hybridizing to complementary sequences produced by the pathogen in the biological fluids. Such techniques, such as PCR, Northern or Southern hybridizations etc. are well known in the art.

It should be understood by one of skill in the art that any number of conventional protein assay formats, particularly immunoassay formats, or nucleic acid assay formats, may be designed to utilize the isolated antigens and antibodies or their nucleic acid sequences or anti-sense sequences of this invention for the detection of *B. burgdorferi* infection in animals and humans. This invention is thus not limited by the selection of the particular assay format, and is believed to encompass assay formats which are known to those of skill in the art.

V. Diagnostic Kits

For convenience, reagents for ELISA or other assays according to this invention may be provided in the form of kits. Such kits are useful for diagnosing infection with *B. burgdorferi* in a human or an animal sample. Such a diagnostic kit contains an antigen of this invention and/or at least one antibody capable of binding an

antigen of this invention, or the nucleic acid sequences encoding them, or their anti-sense sequences. Alternatively, such kits may contain a simple mixture of such antigens or sequences, or means for preparing a simple mixture.

These kits can include microtiter plates to which the IR_{1,5} or IR₆ peptides, proteins, antibodies, or nucleic acid sequences of the invention have been pre-adsorbed, various diluents and buffers, labeled conjugates for the detection of specifically bound antigens or antibodies, or nucleic acids and other signal-generating reagents, such as enzyme substrates, cofactors and chromogens. Other components of these kits can easily be determined by one of skill in the art. Such components may include polyclonal or monoclonal capture antibodies, antigen of this invention, or a cocktail of two or more of the antibodies, purified or semi-purified extracts of these antigens as standards, MAb detector antibodies, an anti-mouse or anti-human antibody with indicator molecule conjugated thereto, an ELISA plate prepared for absorption, indicator charts for colorimetric comparisons, disposable gloves, decontamination instructions, applicator sticks or containers, and a sample preparator cup. Such kits provide a convenient, efficient way for a clinical laboratory to diagnose *B. burgdorferi* infection.

VI. Therapeutic Compositions

The antigens, antibodies, nucleic acid sequences or anti-sense sequences of the invention, alone or in combination with other antigens, antibodies, nucleic acid sequences or anti-sense sequences may further be used in therapeutic compositions and in methods for treating humans and/or animals with Lyme Disease. For example, one such therapeutic composition may be formulated to contain a carrier or diluent and one or more of the antibodies of the invention. Suitable pharmaceutically acceptable carriers facilitate administration of the proteins but are physiologically inert and/or nonharmful.

Carriers may be selected by one of skill in the art. Exemplary carriers include sterile saline, lactose, sucrose, calcium phosphate, gelatin, dextran, agar, pectin, peanut oil, olive oil, sesame oil, and water. Additionally, the carrier or diluent

may include a time delay material, such as glycerol monostearate or glycerol distearate alone or with a wax. In addition, slow release polymer formulations can be used.

Optionally, this composition may also contain conventional
5 pharmaceutical ingredients, such as preservatives, or chemical stabilizers. Suitable ingredients which may be used in a therapeutic composition in conjunction with the antibodies include, for example, casamino acids, sucrose, gelatin, phenol red, N-Z amine, monopotassium diphosphate, lactose, lactalbumin hydrolysate, and dried milk.

Alternatively, or in addition to the antibodies of the invention, other
10 agents useful in treating Lyme disease, e.g., antibiotics or immunostimulatory agents and cytokine regulation elements, are expected to be useful in reducing or eliminating disease symptoms. Agents which can be used to suppress or counteract the immune suppressants released by the tick vector or the spirochete should act to assist the natural immunity of the infected human or animal. Thus, such agents may operate in
15 concert with the therapeutic compositions of this invention. The development of therapeutic compositions containing these agents is within the skill of one in the art in view of the teachings of this invention.

According to the method of the invention, a human or an animal may be treated for Lyme Disease by administering an effective amount of such a therapeutic
20 composition. An "effective amount" may be between about 0.05 to about 1000 $\mu\text{g/mL}$ of an antibody of the invention. A suitable dosage may be about 1.0 mL of such an effective amount. Such a composition may be administered 1 - 3 times per day over a 1 day to 12 week period. However, suitable dosage adjustments may be made by the attending physician or veterinarian depending upon the age, sex, weight
25 and general health of the human or animal patient. Preferably, such a composition is administered parenterally, preferably intramuscularly or subcutaneously. However, it may also be formulated to be administered by any other suitable route, including orally or topically.

VII. Vaccine Compositions

The present invention provides a vaccine composition containing an IR₁₋₅ or IR₆ protein or peptide of the invention or mixtures thereof and a pharmaceutically acceptable carrier or diluent. In another embodiment, the invention provides a vaccine composition containing a nucleic acid sequence of the invention, or mixtures thereof, and a pharmaceutically acceptable carrier or diluent. Combinations of these antigen(s) of this invention with other antigens of *B. burgdorferi*, such as the OspA and OspB proteins, BmpA, B, C or D proteins, or fragments thereof are also encompassed by this invention.

Exemplary carriers are as described above for therapeutic compositions. Optionally, the vaccine composition may further contain adjuvants, preservatives, chemical stabilizers, or other antigenic proteins. Typically, stabilizers, adjuvants, and preservatives are optimized to determine the best formulation for efficacy in the target human or animal. Suitable exemplary preservatives include chlorobutanol, potassium sorbate, sorbic acid, sulfur dioxide, propyl gallate, the parabens, ethyl vanillin, glycerin, phenol, and parachlorophenol.

One or more of the above described vaccine components may be admixed or adsorbed with a conventional adjuvant. The adjuvant is used to attract leukocytes or enhance an immune response. Such adjuvants include, among others, Ribi, mineral oil and water, aluminum hydroxide, Amphigen, Avridine, L121/squalene, D-lactide-poly(lactide)/glycoside, pluronic polyols, muramyl dipeptide, killed *Bordetella*, and saponins, such as Quil A. In addition, a vaccine composition of the invention may further comprise other, non-*B. burgdorferi* antigens, including, *Bordetella bronchiseptica*, canine parvovirus, canine distemper, rabies, *Leptospiridia*, canine coronavirus, and canine adenovirus. Other vaccinal antigens originating from other species may also be included in these compositions, e.g., feline coronavirus, etc.

The invention thus also encompasses a prophylactic method entailing administering to an animal or human an effective amount of such a composition. The vaccine compositions of the invention are administered in an "effective amount", that is, an amount of antigen that is effective in a route of administration to provide a

vaccinal benefit, i.e., protective immunity. Suitable amounts of the antigen can be determined by one of skill in the art based upon the level of immune response desired. In general, however, a protein-based vaccine composition contains between 1 ng to 1000 mg antigen, and more preferably, 0.05 μ g to 1 mg per mL of antigen.

- 5 Generally, a DNA-based vaccine contains a peptide or protein antigen of the invention optionally under the control of regulatory sequences. Where the antigen-encoding DNA is carried in a vector, e.g., a viral vector, a dose may be in the range of 1×10^{-3} pfu to 1×10^{12} pfu.

- Other suitable doses of the vaccine composition of the invention can be readily determined by one of skill in the art. Generally, a suitable dose is between 0.1 to 5 mL of the vaccine composition. In general, the vaccine will be administered once on a seasonal basis. Each tick season, usually in the spring, a booster should be administered. The vaccine may be administered by any suitable route. However, parenteral administration, particularly intramuscular, and subcutaneous, is the preferred route. Also preferred is the oral route of administration. Routes of administration may be combined, if desired, or adjusted. Further, depending upon the human patient or the animal species being treated, i.e. its weight, age, and general health, the dosage can also be determined readily by one of skill in the art.

VIII. Drug Screening and Development

- 20 The proteins, antibodies and polynucleotide sequences of the present invention may also be used in the screening and development of chemical compounds or proteins which have utility as therapeutic drugs or vaccines for the treatment or diagnosis or prevention of Lyme Disease. As one example, a compound capable of binding to IR₁₋₅ or IR₆ and preventing its biological activity may be a useful drug component for the treatment or prevention of Lyme Disease. The methods described herein may also be applied to fragments of IR₁₋₅ or IR₆, and, particularly, to epitopes within these fragments.

Suitable assay methods may be readily determined by one of skill in the art. Where desired, and depending on the assay selected, the selected antigen(s), e.g.,

IR₆, may be immobilized directly or indirectly (e.g., via an anti-IR₆ antibody) on a suitable surface, e.g., in an ELISA format. Such immobilization surfaces are well known. For example, a wettable inert bead may be used. Alternatively, the selected antigen, e.g., IR₆, may be used in screening assays which do not require

5 immobilization, e.g., in the screening of combinatorial libraries. Assays and techniques exist for the screening and development of drugs capable of binding to an antigen of this invention, e.g., IR₆. These include the use of phage display system for expressing the antigenic protein(s), and using a culture of transfected *E. coli* or other microorganism to produce the proteins for binding studies of potential binding

10 compounds. See, for example, the techniques described in G. Cesarini, FEBS Letters, 307(1):66-70 (July 1992); H. Gram *et al.*, J. Immunol. Meth., 161:169-176 (1993); C. Summer *et al.*, Proc. Natl. Acad. Sci., USA, 89:3756-3760 (May 1992), incorporated by reference herein.

Other conventional drug screening techniques may be employed using

15 the proteins, antibodies or polynucleotide sequences of this invention. As one example, a method for identifying compounds which specifically bind to a protein of this invention, e.g., IR₆, can include simply the steps of contacting a selected IR₆ protein with a test compound to permit binding of the test compound to IR₆; and determining the amount of test compound, if any, which is bound to the IR₆ protein.

20 Such a method may involve the incubation of the test compound and the IR₆ protein immobilized on a solid support. Similar methods may be employed for one or more of the cassette string proteins.

Typically, the surface containing the immobilized ligand is permitted to come into contact with a solution containing the protein and binding is measured

25 using an appropriate detection system. Suitable detection systems include the streptavidin horse radish peroxidase conjugate, direct conjugation by a tag, e.g., fluorescein. Other systems are well known to those of skill in the art. This invention is not limited by the detection system used.

Another method of identifying compounds which specifically bind to

30 IR₆ or another peptide or protein of this invention can include the steps of contacting

the peptide or protein, e.g., IR₆, immobilized on a solid support with both a test compound and the protein sequence which is a receptor for IR₆ to permit binding of the receptor to the IR₆ peptide or protein; and determining the amount of the receptor which is bound to the IR₆ peptide. The inhibition of binding of the normal protein by
5 the test compound thereby indicates binding of the test compound to the IR₆ peptide. Similar methods may be employed for one or more of the cassette string proteins.

Thus, through use of such methods, the present invention is anticipated to provide compounds capable of interacting with IR₆ or portions thereof, and/or the IR₁₋₅ peptides or proteins and either enhancing or decreasing the protein's biological
10 activity, as desired. Such compounds are believed to be encompassed by this invention.

The following examples illustrate the preferred methods for obtaining protein antigens of the invention and preparing the assays and compositions of the invention. Significantly, these examples indicate that the IR₁₋₅ and IR₆ peptides and
15 proteins of this invention are useful for diagnosis and prophylaxis against Lyme disease and may improve Lyme serology. These examples are illustrative only and do not limit the scope of the invention.

Example 1 - Materials and Methods for Identification of IR₁₋₅ and IR₆ Peptides

A. *Animals, animal infections and spirochetal strains.*

20 Rhesus monkeys (2 to 4-yr-old, *Macaca mulatta*) were infected by the bite of *Ixodes scapularis* nymphal ticks that were themselves infected either with *B. burgdorferi* sensu stricto strains JD1 or B31. Mice (6 to 8-wk-old C3H/HeN, Jackson Laboratories, Bar Harbor, ME) were infected with *B. burgdorferi* sensu stricto strain Sh-2-82 (low passage, a gift from Denee Thomas, University of Texas
25 Health Science Center, San Antonio, TX) by subcutaneous needle inoculation with 1×10^8 spirochetes administered in 1 ml of BSK-H medium (Sigma Chemical Co., St. Louis, MO), or by the bite of B31-infected *Ixodes scapularis* nymphal ticks. *B. garinii* strain IP90 (low passage) was obtained from the Centers for Disease Control

and Prevention (CDC, Fort Collins, CO). When required, spirochetes were cultivated in BSK-H medium as described previously [Philipp, *Infect. Immun.*, 61:3047-3059 (1993)].

5 B. *Cloning, sequencing and expression of the 7-1 cassette segment of IP90.*

A library of randomly sheared total DNA from *B. garinii* IP90 was constructed in the λ ZAP II bacteriophage vector (Stratagene, La Jolla, CA) following a procedure described previously [R. Ramamoorthy et al, *Infect. Immun.*, 64:1259-1264 (1996)]. The library was screened with a pool of plasma collected from
10 rhesus monkeys within the first 10 weeks after tick inoculation with *B. burgdorferi* JD1. On immunoblots of whole-cell extracts of *B. garinii* IP90 this plasma pool reacted strongly only with three components, namely flagellin, an unidentified 60-kDa protein, and an Ag which was the IP90 homolog of the 34 kDa VlsE of B31 [J.R. Zhang et al, *Infect. Immun.*, 66:3698-3704 (1998)]. After several rounds of screening,
15 eleven clones were rescued into the pBluescript phagemid (Stratagene). The recombinant plasmids were purified and used to transform cells of the SURE strain of *E. coli* (Stratagene). Several transformants were selected from each original clone, the presence of the insert was confirmed, and one such transformant from each clone was grown, induced for expression, lysed and analyzed by immunoblot analysis with the
20 original plasma pool. One of the eleven cloned fragments (named 7-1) hybridized to all others by dot-blot hybridization. This fragment was selected for overexpression and purification on the basis of the strong reactivity of the expressed protein with the plasma Abs. Sequencing of the 7-1 insert was performed by standard procedures [J. Sambrook et al, *Molecular cloning: A Laboratory Manual*, Cold Spring Harbor
25 Laboratory, Cold Spring Harbor, NY (1989)] after generating *Bal* 31 nested deletions and subcloning the partially deleted fragments. Ab from the original plasma pool that was affinity purified using the recombinant protein (P7-1) expressed by clone 7-1 as immunoabsorbant reacted with the putative IP90 VlsE on immunoblot of *B. garinii* lysates. The 7-1 insert was subcloned into the pQE expression system for

overexpression and purification of the polypeptide (Qiagen Inc., Chatsworth, CA). The protein concentration was determined using the Bio-Rad protein assay kit (Bio-Rad Laboratories, Richmond, CA) with BSA as a standard.

C. *P7-1 ELISA.*

5 This protocol was initially used to monitor monkey Ab responses to P7-1. Ninety-six-well ELISA plates (Corning Inc., Corning, NY) were coated with 100 μ l per well of a solution of P7-1 at 0.1 μ g/ml in coating buffer (0.1 M carbonate buffer, pH 9.2) at 4°C overnight. Plates were blocked with 200 μ l per well of 5% FCS in PBS/T (10 mM sodium phosphate, 150 mM NaCl and 0.1% Tween-20, pH 7.4) for 1 hour at room temperature. After washing 3 times with
10 PBS/T, 50 μ l per well of serum samples from *B. burgdorferi*-infected monkeys diluted 1/100 in 5% FCS in PBS/T were incubated for 1 hour at 37°C. Plates were washed and incubated with 50 μ l of (1) a mixture of biotinylated goat anti-human IgG (γ -chain specific) and IgM (μ -chain specific) Abs at the dilution recommended by the
15 manufacturer (Vector Laboratories, Inc., Burlingame, CA) for 1 hour at 37°C, (2) avidin-horseradish peroxidase complex also at the dilution recommended by the manufacturer (Vector) for 30 minutes at 37°C, (3) a solution containing 2 g/l ortho-phenylenediamine and 0.03% hydrogen peroxide (both from Sigma) in 0.1 M citrate-sodium-phosphate buffer pH 5.0 for 10 minutes at room temperature. The
20 reaction was stopped with 50 μ l of 4 NH_2SO_4 . OD at 490 nm was determined. Goat anti-human IgG and IgM fully crossreacted with monkey Ab.

D. *Identification of conserved sequences and prediction of antigenicity of invariable regions of the VlsE variable domain.*

The deduced amino acid sequence of P7-1 was compared with
25 sequences available in the GenBank data base (National Center for Biotechnology Information, Rockville, MD) using the BlastP algorithm [J. Zhang and T.L. Madden, Genome Res., 6:649-656 (1997)]. The antigenicity of the entire P7-1 polypeptide was analyzed using the Hopp-Woods scale [T.P. Hopp and K. R. Woods, Proc. Natl. Acad. Sci. USA, 78:3824-3828 (1981)], and identities of invariable regions of P7-1
30 with homologous cassette segments from strains B31 and 297 were calculated after

aligning the cassette segments using the pam250 algorithm, and MacVector 5.0 computer software (Eastman Kodak Company, New Haven, CT).

E. Peptide synthesis and conjugation to biotin.

A 26-mer peptide (C_6) which reproduced the IR_6 sequence of
5 VlsE cloned from *B. garinii* strain IP90 was prepared using the
fluorenylmethoxycarbonyl synthesis protocol [G. Barony and R.B. Merrifield, *The Peptides: Analysis, Synthesis, & Biology*, Academic Press, pp. 3-285 (1980)]. A
cysteine residue was included at the N-terminus and used as biotinylation site.
Biotinylation was performed by the N-succinimidyl maleimide carboxylate method.
10 The maleimide reagent was from Molecular Probes (Eugene, OR) and the protocol
suggested by the manufacturer was followed.

F. Human serum samples.

A panel of 41 human serum samples was kindly provided by the
CDC. All the samples were collected from Lyme disease patients who had signs and
15 symptoms that satisfied the CDC clinical case definition [Morbidity and Mortality
Weekly Report, 39, No. RR-13, 19-20 (1990)]. Four serum samples from chronic
Lyme disease patients were obtained from the National Institutes of Health.
Ninety-seven serum samples obtained from hospitalized patients in an area not
endemic for Lyme disease were used as negative controls.

20 *G. C_6 Peptide ELISA of the Invention.*

Ninety-six-well ELISA plates were coated with 100 μ l per well
of 4 μ g/ml streptavidin (Pierce Chemical Company, Rockford, IL) in coating buffer
and incubated at 4°C overnight. The remaining steps were conducted in a rotary
shaker at room temperature. After two 3-minute washes with 200 μ l per well of
25 PBS/T at 200 rpm, 200 μ l of 5 μ g/ml biotinylated C_6 peptide dissolved in blocking
solution (PBS/T supplemented with 5% nonfat dry milk [Carnation, Nestle Food
Company, Glendale, CA]) was applied to each well. The plate was shaken at 150 rpm
for 2 hours. After three washes with PBS/T as above, 50 μ l of serum (mouse, monkey
or human) diluted 1:200 with blocking solution was added to each well. The plate was
30 incubated at 150 rpm for 1 hour and then washed three times with PBS/T. Each well

then received 100 μ l of 0.2 μ g/ml goat anti-monkey IgG (γ -chain specific, [Kirkegaard & Perry Laboratories, Gaithersburg, MD]), 0.5 μ g/ml anti-mouse IgG (heavy and light-chain specific, [Sigma]), or 0.1 μ g/ml anti-human IgG (heavy and light-chain specific, [Pierce]), each conjugated to horseradish peroxidase and dissolved in blocking solution. The plate was incubated for 1 hour while shaking. After four washes with PBS/T each for 3, 4, 5 and 6 minutes, respectively, the Ag-Ab reaction was probed using 100 μ l of a solution composed of the chromogen 3,3',5,5'-tetramethylbenzidine at 0.2 mg/ml and 0.01% hydrogen peroxide in the buffer supplied by the manufacturer [TMB Microwell Peroxidase Substrate System, Kirkegaard & Perry], and color was allowed to develop for 10 minutes. The enzyme reaction was stopped by addition of 100 μ l of 1 M H_3PO_4 . OD was measured at 450 nm with an ELISA plate spectrophotometer model SLT Spectra with the Soft2000 Software (SLT Lab Instruments, Germany).

H. *Competitive ELISA.*

Ninety-six-well ELISA plates were coated with 100 μ l per well of 0.3 μ g/ml of purified recombinant P7-1 dissolved in coating buffer at 4°C overnight. After blocking with blocking solution, 25 μ l of this solution with 0, 1, 4, 16, 64, 256 or 1024 ng of the C_6 peptide was added to each well. A 25- μ l volume of a 1:100 dilution of the appropriate serum in blocking solution also was added to each well. The remaining steps were performed as related in paragraph G above describing the C_6 peptide ELISA.

I. *Preparation of rabbit anti- C_6 peptide antiserum.*

The C_6 peptide was covalently linked to keyhole limpet hemocyanin (KLH) by the N-succinimidyl maleimide carboxylate method. The maleimide reagent was from Molecular Probes (Eugene) and the protocol suggested by the manufacturer was followed. Six-month-old New Zealand White rabbits were given three injections at bi-weekly intervals of 200 μ g of conjugated Ag emulsified with Freund's complete (first injection) or incomplete adjuvant (remaining injections). Ten days after the last injection the Ab titer was determined by the peptide ELISA and immunoblot analysis using IP90 spirochete whole-cell lysates as Ag.

J. *Immunoprecipitation and immunoblot.*

Immunoprecipitation was conducted at 4°C. Approximately 1.5 x 10¹⁰ IP90 spirochetes harvested at stationary growth phase were extracted in 4.5 ml solubilization buffer (50 mM Tris-HCl, 1% Triton X-100, 1 mM EDTA, pH 7.6) for 30 minutes. The mixture was centrifuged at 13,000 x g for 30 minutes and the supernatant was collected. Each of 1.5 ml of this supernatant was mixed with 30 µl of preimmune or immune rabbit serum and incubated for 30 minutes. Fifty microliters of drained ImmunoPure™ Immobilized Protein G (Pierce) preequilibrated in solubilization buffer was then added and allowed to incubate for an additional 30 minutes. After washing the gel twice with excess volumes of this buffer by centrifugation at 3,000 x g for 20 minutes, 150 µl of nonreducing SDS-PAGE sample buffer (125 mM Tris-HCl, 3% SDS and 20% glycerol, pH 6.8) was added. The suspension was incubated at room temperature for 30 minutes and then centrifuged at 16,000 x g for 30 minutes. Ten microliters of supernatant was loaded onto each of ten lanes of a SDS 12% polyacrylamide mini-gel. Separated proteins were electrotransferred to nitrocellulose in Towbin transfer buffer. After incubating in blocking solution for 2 hours, the blot was incubated for 1 hour in rabbit anti-C₆ serum diluted 1:2,000 with blocking solution. After 3 washes with PBS/T, the blot was incubated in blocking solution with 0.5 µg/ml goat anti-rabbit IgG conjugated to horseradish peroxidase (Pierce) for 1 hour. The Ag-Ab reaction was probed in PBS/T supplemented with 0.05% 4-chloro-naphthol (Sigma), 0.015% hydrogen peroxide and 17% methanol.

K. *Direct Immunofluorescence.*

Rabbit IgG was purified from preimmune or immune serum using conventional ammonium sulfate precipitation. The purity and concentration of IgG preparations were assessed using SDS-PAGE and the Bio-Rad protein assay kit, respectively. Purified IgG was conjugated to FITC as per the manufacturer's instruction (Pierce). For labeling, spirochetes were either unfixed or acetone fixed. For the fluorescent labeling of unfixed spirochetes, approximately 10 spirochetes that were harvested from 1.0 ml of IP90 culture grown to stationary phase by

centrifugation at 4,000 x g for 20 minutes were gently resuspended in 100 µl PBS supplemented with 2 µg rabbit anti-C₆ IgG-FITC conjugate, and incubated for 1 hour. After 2 washes with excess volumes of PBS by centrifugation at 16,000 x g for 5 minutes, spirochetes were resuspended in 100 µl PBS, applied to microscope slides and counted under both darkfield and fluorescent microscopes. The ratio of fluorescent to total spirochetes was thus calculated. The same procedure was performed on acetone fixed spirochetes. For this purpose, organisms harvested from 1.0 ml culture fluid were suspended in 1.5 ml acetone, incubated for 20 minutes, and then centrifuged at 16,000 x g for 5 minutes. After one wash with PBS, the fixed spirochetes were stained with fluorescent anti-C₆ Ab as described for unfixed spirochetes. Goat anti-*B. burgdorferi* Ab FITC conjugate (Kirkegaard & Perry Laboratories) was used as positive control.

L. ADCK assay.

As a source of complement, serum samples were collected from normal rhesus macaques, pooled and stored in small aliquots at -70°C until used. Serum chosen for this purpose did not contain cross-reactive anti-*B. burgdorferi* Abs as determined by immunoblot analysis using whole cell lysates of *B. burgdorferi* as Ag. To perform the ADCK assay, spirochetes were cultured in BSK-H medium until they reached mid-logarithmic phase (about 2×10^7 cells per ml). A total of approximately 5×10^5 spirochetes in 25 µl of BSK-H medium was added to each well of a 96-well plate (Corning Inc.). A volume of 50 µl of heat-inactivated (56°C, 30 minutes) serum sample appropriately diluted in the same medium was already dispensed in each well. The plate was incubated at 34°C for 30 minutes before the addition of 25 µl of complement preparation (normal monkey serum). After 24 hours of incubation at 34°C in a humidified atmosphere of 3% CO₂, 5% O₂ and the balance of N₂, 5 µl of each sample was removed, and dead (nonmotile) and live (motile) spirochetes were counted under a darkfield microscope. Monkey anti-OspA antiserum was used as positive control [J.M. Nowling and M.T. Phillip, *Infect. Immun.*, 67:443-445 (1999)].

Example 2 - The invariable regions of the VlsE variable domain are conserved among strains and genospecies of *B. burgdorferi*.

The Ab response to the recombinant P7-1 polypeptide in monkeys that had been tick inoculated with either B31 or JD1 *B. burgdorferi* spirochetes was assessed and it was noted that the response was detectable within the first three weeks post-infection (PI), and that it persisted at least until week 10 PI, the longest time point measured in this initial experiment (Fig. 1). The deduced amino acid sequence of the *B. garinii* P7-1 recombinant polypeptide is shown in Fig. 2. It is depicted aligned with sequences from VlsE cassette segments of *B. burgdorferi* strains B31 [J.R. Zhang et al, Cell, 89:275-285 (1997)] and 297 [H. Kawabata et al, Microb. Pathog., 24:155-166 (1998)]. In B31, the cassette segment is comprised between the repeats EGAIKG [SEQ ID NO: 2] (Fig. 2) [Zhang, cited above]. The six variable regions of the B31 VlsE cassette segment [Zhang, cited above] are doubly underlined. The invariable regions IR₁₋₆ of the IP90 VlsE variable domain (Fig. 2) are clearly conserved across both genospecies and strains of *B. burgdorferi* sensu lato. With the exception of IR₁, which is 78% conserved in strain B31 and only 11% in 297, all other invariable regions are between 80 and 90% conserved with respect to the P7-1 VlsE cassette segment of IP90 (Table 1).

Table 1

Identity between invariable regions from VlsE cassette segments of *B. burgdorferi* strains B31 and 297 and *B. garinii* IP90 (P7-1).

Invariable Region	Percent Identity (%)		
	B31 vs. IP90	297 vs. IP90	Median
IR ₁	78	11	45
IR ₂	81	88	85
IR ₃	86	86	86
IR ₄	80	80	80
IR ₅	80	90	85
IR ₆	85	88	87

The antigenicity of the invariable regions of P7-1 was assessed with the Hopp-Woods hydrophilicity algorithm [Proc. Natl. Acad. Sci. USA, 78:3824-3828 (1981)]. Reputedly, this method has been more successful than similar ones for identifying protein antigenic determinants [T.P. Hopp, Pept. Res., 6:183-190 (1993)].

5 Overall, the invariable regions of the IP90 variable domain showed negative hydrophilicity values, possibly indicative of their lack of exposure on the surface of the protein. Sequences with positive values were composed of fewer than 6 amino acids and thus not likely to configure antigenic sites. In contrast, IR₆ contained six or more contiguous amino acids with relatively high positive hydrophilicity values (1.7).

10 IR₆ was also slightly more conserved than other invariable regions, with a median identity of 87% when using the IP90 sequence as the reference strain (Table 1).

Example 3 - Experimental assessment of the antigenicity of IR₆.

To evaluate experimentally the antigenicity of IR₆, the C₆ peptide, whose primary structure encompassed that of IR₆ with an added N-terminal cysteine residue for biotinylation, was synthesized as described in Example 1. The resulting C₆

15 peptide has the amino acid sequence: CMKKDDQIAAAMVLRGMAKDGQFALK [SEQ ID NO: 8].

Serum samples from bleeds obtained from 10 rhesus macaques that were infected either with JD1 (6 animals) or B31 spirochetes were used to examine

20 the antigenicity of IR₆ in monkeys. Anti-C₆ ELISA Ab levels shown in Fig. 3A were present in serum samples at 4-6 weeks PI and remained as high or higher for up to three years PI (data not shown).

Mice also responded vigorously to this region. Serum samples collected 4-6 weeks PI from 10 mice infected either with *B. burgdorferi* strain Sh-2-82 (6

25 animals) or B31 showed high levels of anti-C₆ Abs (Fig. 3B). This result further confirmed the antigenicity of IR₆ and reaffirmed this region's antigenic conservation.

In humans, the antigenicity of IR₆ was examined with the aid of the CDC panel of 41 serum samples. Ab directed to C₆ was found in 36 of the 41 samples (Table 2). N: negative result; P: positive result.

Table 2

Antigenicity of IR₆ in human patients whose signs and symptoms satisfied the CDC Lyme disease case definition.

	Serum ID#	C ₆ ELISA OD	Serum ID#	C ₆ ELISA OD
5	90-2111	P 3.055	91-1352	P 1.763
	90-2436	P 2.541	91-1353	P 0.550
	90-2622	P 3.099	91-1354	P 1.140
	90-2631	N 0.276	91-1841	P 0.607
	90-2668	P 1.625	91-1458	P 1.056
10	91-0521	P 1.423	91-1842	P 1.423
	91-0531	P 3.330	91-1843	P 1.733
	91-0532	P 1.942	91-1844	P 1.879
	91-0533	P 2.178	91-1845	N 0.222
	91-0544	P 3.091	91-1846	N 0.338
15	91-0794	P 0.600	91-1847	P 1.676
	91-0865	P 3.274	92-0057	P 1.735
	91-0900	P 2.386	92-1682	N 0.404
	91-0943	P 2.959	92-1941	N 0.432
	91-1104	P 1.642	92-1982	P 0.832
20	91-1222	P 2.274	93-0206	P 0.633
	91-1347	P 0.960	93-0208	P 0.600
	91-1348	P 3.233	93-1414	P 1.025
	91-1349	N 0.448	93-1426	P 0.832
	91-1350	P 1.246	94-0357	P 0.739
25	91-1351	P 0.843		

The result of this experiment also underscores both the antigenicity and antigenic conservation of IR₆, as the CDC serum panel was composed of samples collected from several Lyme disease endemic areas in the United States and must therefore encompass Abs to multiple strains of *B. burgdorferi*.

5 Example 4 - IR₆ is the only immunodominant invariable region in both monkeys and humans

In addition to IR₆, the VlsE variable domain contains five other invariable regions (Fig. 2). To determine if any of these regions is antigenic, a competitive ELISA was performed, using P7-1 as Ag attached to an ELISA plate, in
10 the presence of increasing concentrations of the C₆ peptide. P7-1 includes the invariable regions IR₁₋₆ (Fig. 2). Ten serum samples from infected monkeys, 4 human serum samples randomly selected from the CDC panel, and an additional 4 serum samples from chronic Lyme disease patients provided by the National Institutes of Health were tested. Representative results for 4 human and 4 monkey serum samples
15 are presented in Fig. 4. Addition of C₆ almost completely inhibited the reaction of monkey and human serum Abs reactive with P7-1. Thus IR₆ appears to be the only immunodominant invariable region within the VlsE variable domain in both monkeys and humans. When the competitive ELISA was used to analyze infected mouse serum, the maximum inhibition resulting from the addition of C₆ was no larger than
20 40%, indicating that other invariable regions may be antigenic in mice (data not shown).

Example 5 - Exposure of IR₆ on the surface of VlsE.

The exposure of IR₆ on the surface of the VlsE protein was examined by immunoprecipitation with the rabbit anti-C₆ antiserum. VlsE from *B. garinii* strain
25 IP90 spirochetes was extracted in solubilization buffer and immunoprecipitated with protein-G-agarose in the presence of either rabbit anti-C₆ antiserum or serum obtained from the same rabbit prior to immunization. Presence of VlsE in the

immunoprecipitates was then assessed on immunoblots reacted with the anti-C₆ antiserum. The VlsE of IP90 was immunoprecipitable with the anti-C₆ antiserum but not with normal serum. This result indicates that IR₆ is exposed on the VlsE surface.

Example 6 - IR₆ is not accessible to Ab on the outer membrane of the spirochete.

5 Exposure of IR₆ on the spirochetal surface was assessed by immunofluorescence. While FITC-conjugated rabbit anti-C₆ Ab extensively labeled all of the acetone-fixed IP90 spirochetes, it labeled few (less than 5%) of the unfixed spirochetes. The unfixed spirochetes that were labeled exhibited a discontinuous fluorescent pattern, whereas fixed spirochetes fluoresced uniformly. Control anti-*B.*
10 *burgdorferi* Ab-FITC conjugate labeled both fixed and unfixed spirochetes. These results indicate that IR₆ is constrained from the surface of the spirochete although the VlsE is surface-exposed [Zhang, *Cell*, 89:275-285 (1997)]. This is consistent with the observation made in the ADCK assay, in which anti-C₆ antiserum had no significant killing activity compared with preimmune serum although monkey anti-OspA
15 antiserum killed all spirochetes in the same experiment (data not shown).

Example 7 - IR₁₋₅ and IR₆

This invention demonstrates that the amino acid sequences of the six invariable regions described previously in the cassette segments of VlsE [Zhang, cited above] are conserved, at least in two *Borrelia* genospecies, *B. garinii* and *B.*
20 *burgdorferi* sensu stricto. Further, it has been illustrated with data from the literature that such conservation is also retained among strains of the latter genospecies. This is evident from the high level of identity between the deduced amino acid sequence of the invariable regions within the variable domain of IP90 (*B. garinii*), which was cloned, sequenced and identified, and those of the B31 and 297 cassette segments
25 (Fig. 2, Table 1).

The sequence conservation of the six invariable regions across strain and genospecies barriers indicates that these regions are important in the role VlsE plays in the physiology of *B. burgdorferi*. One would therefore expect that such

sequences are not antigenic in hosts with a chronic *B. burgdorferi* infection or would be otherwise inaccessible to Ab, either because they are conformationally buried within the VlsE molecule or unavailable on the spirochetal surface. The Hopp-Woods algorithm indicated that, with the exception of IR₆, all of the other invariable regions
5 were either not antigenic or had very low antigenicity.

The predicted antigenicity of IR₆ was confirmed in humans, monkeys and mice (Fig. 3, Table 2). Sera from all of these hosts reacted with the C₆ peptide early and persistently in the course of infection, thus indicating that IR₆ contains one or more epitopes that may be broadly antigenic, regardless of host species. The
10 antigenicity of C₆ was not only manifest independently of host species but also regardless of whether the animals had been infected with the JD1, B31 or Sh-2-82 strains of *B. burgdorferi sensu stricto*. In addition, 36 of 41 human serum samples collected in the Northeast and Midwest of the U. S. from patients with acute or chronic Lyme disease also reacted with this peptide. The 5 serum samples that had no
15 detectable anti-C₆ Ab were obtained from patients who were in early stages of infection. Hence, the negative results may reflect presence of very low serum Ab titers rather than absence of crossreactivity. The C₆ peptide may thus serve as a global diagnostic probe.

The absence of, or relatively low, antigenicity of IR₁₋₅ was underscored
20 by the results of the competition experiments (Fig. 4). The fact that the binding of most of the human and monkey Ab that reacted with P7-1 could be inhibited simply by adding an excess of the C₆ peptide indicates that in these hosts IR₆ is possibly the only immunodominant invariable region within the VlsE variable domain. In mice the results were different, as an added excess of the C₆ peptide was unable to fully inhibit
25 the P7-1 reactivity of mouse anti-Sh-2-82 or anti-B31 Abs. This result implies that some of the other invariable regions may be antigenic in mice. The immunodominance of IR₆ was further underscored by the long-term persistence of anti-C₆ Abs in infected monkeys and in patients with chronic Lyme disease (data not shown).

The Hopp-Woods algorithm also predicted, as expected, that some
30 variable regions of the cassette segment are strongly antigenic (data not shown).

Thus, at least theoretically, the VlsE variable regions could satisfy the antigenic variation paradigm that variable domains are immunodominant. In the case of VlsE, however, variable region immunodominance of individual VlsE molecules may be difficult to achieve, for the recombination process that underlies the mechanism of VlsE variation occurs very rapidly [J.R. Zhang et al, *Infect. Immun.*, 66:3869-3697)].
In spite of strong antigenicity, no variable region might at any time be represented on a large enough number of bacterial cells, or otherwise be expressed long enough, to become immunodominant. Indeed, within the first 4 days PI, as many as 11 predicted amino acid changes per VlsE variant can occur during a *B. burgdorferi* B31 infection in C3H/HeN mice [Zhang, cited above], and unlike with *T. brucei*, *N. gonorrhoeae* and *B. hermsii*, VlsE serotypes of *B. burgdorferi* have never been found.

The finding that IR₆ is exposed on the VlsE surface but not on the spirochetal surface is in agreement with the antigenicity and immunodominance of this region. Features of protein domains such as surface accessibility, hydrophilicity, flexibility and proximity to a site recognized by helper T cells are all important in positively determining domain antigenicity [J.A. Berzofsky and I.J. Berkower, Immunogenicity and antigen structure, in "Fundamental Immunology", William E. Paul, Editor, Raven Press, Ltd., New York, 235-282 (1993)]. While it is possible that solubilization with Triton X-100 may have partially denatured VlsE and thus artificially exposed IR₆, the result of the immune precipitation experiment indicates that IR₆ is accessible on the surface of native VlsE. It is also hydrophilic, as assessed by the Hopps-Wood algorithm. The molecular surface exposure of IR₆ was also indicated by the fact that a small proportion of unfixed spirochetes fluoresced when incubated with FITC-labeled anti-C₆ Ab.

Most of the unfixed spirochetes failed to label with the FITC-conjugated anti-C₆ Ab. This indicates that IR₆ is not exposed on the spirochetal surface. The small proportion of unfixed spirochetes that bound anti-C₆ Ab likely had some degree of membrane damage. Although our procedure for labeling of unfixed spirochetes entailed very gentle manipulations, it is possible that portions of the VlsE molecule not normally exposed on the spirochete surface were nonetheless uncovered

in a fraction of the spirochetes. The fragility of the outer membrane of *B. burgdorferi* has been noted by other investigators [D.L. Cox et al, Proc. Natl. Acad. Sci. USA, 93:7973-7978 (1996)]. Loss of the lp28-1 plasmid, which encodes VlsE, during *in vitro* cultivation [Zhang, Cell, 89:275-285 (1997)], also could explain a negative labeling result. However, the fact that nearly 100% of acetone-fixed spirochetes from the same culture were labeled allows us to rule out this possibility. Acetone fixation probably exposed regions of the VlsE not accessible to Ab on an intact spirochete.

The result of the ADCK experiment was entirely consistent with the immunofluorescent observations, as absence of killing is most likely due to failure of the anti-C₆ Ab to bind to the spirochetal surface. Complement dependent killing of *B. burgdorferi* is facilitated by anti-surface Ab binding and does not depend on the C-activating properties of the Ab. *B. burgdorferi* spirochetes are able to activate complement through an Ab-independent mechanism [S. M. Kochi et al, Infect. Immun., 61:2532-2536 (1993)]. Taken together, the immunofluorescence and ADCK results indicate that IR₆ is cryptic on the spirochetal surface.

While the variable regions of Ags such as the VSG, Vmp or pilin are extremely antigenic, no strong antigenicity of these molecules' invariable regions or their longer invariable domains has been reported [D.M. Reinitz et al, Mol. Biochem. Parasitol., 51:119-132 (1992); K. T. Forest et al, Infect. Immun., 64:644-652 (1996)]. The principal role ascribed to both invariable regions and domains has been the preservation of functional molecular conformations [Reinitz, cited above; Forest, cited above]. It has been hypothesized that chronic host exposure to immunodominant Ags or epitopes diverts the immune system from responding to less antigenic but functionally important Ags or epitopes, thus serving as a protective strategy for persistent pathogens [P. Marrack and J. Kappler, Cell, 76:323-332 (1994)]. Recently, it was demonstrated that when the immunodominant V3 loop epitope of glycoprotein 120 (gp120) of human immunodeficiency virus-1 is masked through site-directed targeting of N-linked glycosylation, the dominant, type-specific neutralizing Ab response is shifted away from V3 to epitopes in the first variable domain (V1) of

gp120 [R.R. Garrity et al, J. Immunol., 159:279-289 (1997)]. Ab responses to conserved domains of gp120 also were observed [Garrity et al, cited above].

The role of the antigenicity and immunodominance of IR₆ is to act as a decoy epitope and contribute to subvert the Ab response to *B. burgdorferi*. Variable regions of VlsE do not, or not always, become immunodominant. The VlsE antigenic variation may serve to inhibit the formation of high avidity Abs to the variable VlsE regions, but not to divert the response away from invariable VlsE regions and invariable domains or other less antigenic but functionally neutralizing *B. burgdorferi* Ag. Only about half of the length of the mature VlsE protein is variable, compared to more than two thirds of proteins such as VSG, pilin or VMP. Moreover, more than half of the variable domain of VlsE is encompassed by invariable regions including the highly immunodominant IR₆. Conserved regions of VlsE other than IR₆ thus may be exposed, per force, on the spirochete surface. IR₆ serves as the decoy epitope for such domains, by suppressing protective immune responses through mechanisms such as clonal restriction and/or idiotypic dysregulation, as has been invoked for HIV-1's V3 [H. Kohler et al, J. Acquired Immune Defic. Syndr., 5:1158-1168 (1993); R. Metals and V. Veljkovic, Vaccine, 13:355-359 (1995)].

Example 8 - An IR₆-Based ELISA for Diagnosis of Lyme Disease

Diagnostic specificity and sensitivity of the C6 peptide ELISA described in Example 1G was examined, as described in this example. This data demonstrates that the ELISA is a simple, sensitive, specific and precise assay which alleviates some of the remaining problems in Lyme disease serodiagnosis. Its synthetic peptide makes it inexpensive to manufacture and makes it useful for diagnosis even in OspA-vaccinated subjects.

As used herein, Sensitivity is defined as True Positives/True Positives + False Negatives, Specificity as True Negatives/True Negatives + False Positives, precision as frequency of obtaining the same result in duplicate analysis of a set of positive and negative specimens. Early localized, early disseminated and late Lyme disease are well defined in A. Steene, N. Engl. J. Med., 321:586-596 (1989).

Positive Predictive Value as True Positives/True Positives + False Positives, and
Negative Predictive Value as True Negatives/True Negatives + False Negatives.

A. *Serum Sources and Peptide ELISA*

1. *Monkey serum samples*

5 Rhesus monkeys (2 to 4-year-old, *Macaca mulatta*)
were infected by the bite of *Ixodes scapularis* nymphal ticks (9 animals), or by needle
inoculation (1 animal) as previously described [Philipp et al., *Infect. Immun.*, 61:3047-
3059 (1993)]. Ticks feed on the animals were themselves infected with spirochetes
of either the JD1 (Philipp et al, 1993, cited above) or B31 [Philipp et al, *Vaccine*,
10 15:1872-1887 (1997)] strains of *B. burgdorferi sensu stricto*. The needle-inoculated
animal received JD1 spirochetes. Blood specimens were collected every one or two
weeks postinoculation and serum samples were stored at -20°C until the peptide
ELISA was conducted. To assess crossreactivity of the C₆ peptide with anti-OspA
antibodies, serum samples from animals that had been administered an OspA vaccine
15 were utilized [Philipp et al, cited above, (1997)]. The vaccine formulation and
administration protocol, which also had been used in human trials [C. Van Hoeske et
al, *Vaccine*, 14:1620-1626 (1996)] have been described. previously [Philipp et al,
cited above, (1997)].

2. *Human serum samples*

20 Human serum samples were obtained from a variety of
sources. To assess diagnostic sensitivity, three serum panels from Lyme disease
patients were used. One was from the Centers for Disease Control and Prevention,
kindly provided by Dr. Martin Schriefer; a second panel was from patients of
Tufts-New England Medical Center; a third panel was from the National Institutes of
25 Health. The CDC serum panel was composed of 40 samples from patients who were
in the convalescent phase of Lyme disease. Twenty-seven of these samples were also
culture confirmed. All of the serum specimens in this panel were from patients whose
case satisfied the CDC Lyme disease case definition [Centers for Disease Control and
Prevention, *Morb. Mort. Wkly Rep.*, 46:20-21 (1997)]. In addition, all of the
30 samples had been tested at the CDC with commercially available kits: ELISA was

performed with Lyme Screen II (bioMerieux, St. Louis, MO) and IgG and IgM with Marblot (MarDx, Carlsbad, CA). The panel from Tufts-New England Medical Center was composed of 157 specimens, of which 39 were from patients in the acute phase of Lyme disease. These patients were in the early (localized or disseminated) phase of Lyme disease. Also in this panel were 39 specimens from individuals in the convalescent phase, 20 from patients who had presented with signs and/or symptoms of early disseminated phase who had presented with signs and/or symptoms of early neuroborreliosis, and 59 from patients with late Lyme disease (49 with Lyme arthritis, and 10 with late neuroborreliosis). All of the patients in the Tufts panel met clinical criteria for Lyme disease diagnosis, as described previously [F. Dressler et al, J. Infect. Dis., 167:392-400 (1993)]. The NIH panel was composed of 13 specimens obtained from patients with post-treatment Lyme disease, defined as persistent or intermittent symptoms for at least 6 months after appropriate antibiotic therapy for Lyme disease. Usual symptoms include widespread musculoskeletal pain and fatigue; memory and/or concentration impairment, radicular pain, paresthesia or dysesthesia. The beginning of the symptoms coincides with, or occurs within 6 months of, the initial *B. burgdorferi* infection. Symptoms are significant enough to interfere with daily life activities, and other causes have been excluded. All of these samples met CDC criteria for seropositivity [US Department of Health and Human Services, Centers for Disease Control and Prevention, Morb. Mortal. Wkly Rep., 44:590-591 (1995)].

To examine diagnostic specificity of the peptide ELISA, a panel of 56 serum specimens was obtained from patients with autoimmune or neurological diseases, spirochetal diseases other than Lyme borreliosis, or other chronic infections was obtained from the NIH. Sera were from patients with multiple sclerosis (n=10), positive anticardiolipin antibody (n=10), positive rheumatoid factor (n=10), positive rapid plasma reagin (n=10), positive antinuclear antibody (n=10), Guillain-Barré syndrome (n=1) or mycobacterial infection (n=5). A second panel, obtained from the CDC, was composed of 9 serum samples from patients with relapsing fever. A third panel of 12 additional syphilis serum samples, (9 from patients with early latent and 3

with late latent disease) was obtained from the syphilis serum bank maintained at the University of California at Los Angeles (UCLA); each serum sample was positive by the *Treponema pallidum* immobilization test. Finally, a panel of 99 specimens was obtained blindly from hospital patients in Louisiana, where Lyme disease is not known to be endemic.

3. *Peptide ELISA*

The C₆ peptide was synthesized as described in Example 1 above. The peptide ELISA was performed as described in Example 1 above. The cut-off OD (0.500) value was defined as the mean OD + 3 standard deviations of 97 serum samples collected from patients of a Louisiana hospital (where Lyme disease is not endemic).

B. *Early and persisting IgG response to C₆ in infected rhesus monkeys*

Serum samples serially collected from ten monkeys that had been inoculated either with the B31 or the JD1 strains of *B. burgdorferi* were tested by ELISA for antibody responses to C₆. IgG antibody was detectable in 7 animals as early as 3 weeks postinoculation. At week 5 postinoculation 9 animals had responded, and all at week 6. Antibody to C₆ remained at high levels in all animals during the entire study period, which was between 25 and 160 weeks. Only the serum samples from JD1-infected monkeys are shown. The IgM anti-C₆ antibody responses were detectable only in some of the animals and did not appear earlier than the corresponding IgG responses.

C. *Sensitivity of the C₆ ELISA*

Forty serum samples obtained by the CDC from patients in the convalescent phase of Lyme borreliosis were assessed with the C₆ ELISA.

Table 3
Sensitivity of C₆ ELISA

Panel #	Infection Course	Sample No.	Positive	Negative	Sensitivity (%)
1 (CDC)	Convalescent	40	34	6	85
2	Acute	39	29	10	74
	Convalescent	39	35	4	90
	Early Dissem. (neuroborreliosis)	20	19	1	95
	Late (arthritis)	49	49	0	100
	Late (neuroborreliosis)	10	10	0	100
3	Chronic	19	13	6	68
Overall		218	190	28	90

Thirty-four were positive (Table 3), thus yielding a sensitivity of detection of 85%. An assessment of the same samples performed at the CDC yielded a sensitivity of 80% (32/40) with a conventional ELISA and 75% (30/40) with the combination of both IgG and IgM immunoblots.

One hundred and fifty-nine additional serum samples collected from patients with different phases of Lyme disease at Tufts-New England Medical Center also were assessed with the C₆ ELISA. The sensitivity was 74% (29/39) for patients in the acute disease phase, 90% (35/39) for convalescent-phase patients, 95% (19/20) for early disseminated (neuroborreliosis), and 100% (59/59) for late Lyme disease patients, respectively (Table 3). Of these, 49 had late Lyme arthritis and 10 had late neuroborreliosis (Table 3). All of the "acute" patients in this panel had signs of localized infection (erythema migrans) and some had, in addition, signs or

symptoms of disseminated infection. An analysis of the anti-C₆-antibody-positive fraction of serum samples taken at consecutive weeks after the onset of signs/symptoms showed that 80-90% of the samples taken at 3 weeks or later after disease onset were positive. An additional test of 13 serum specimens collected from patients with chronic Lyme disease yielded a sensitivity of 62% (8/13) (Table 3).

Acute and convalescent phase serum specimens from Panel 2 were employed to examine the IgM antibody response to C₆. Of the 39 samples in each category, none were only IgM antibody positive, 16 "acute" and 10 "convalescent" specimens were both IgG and IgM antibody positive, and 13 and 25 were only IgG antibody positive in the acute and convalescent categories, respectively.

The sensitivity of the C₆ ELISA ranged from 62% to 100% depending on when in the course of infection the serum samples were collected, and on the clinical definition of the patients' signs and symptoms (Table 3). Based on the results of our longitudinal analysis of the anti-C₆ antibody response in infected rhesus monkeys, it is unlikely that such a response may be detectable with good sensitivity earlier than one to two weeks postinfection. However, by week 5 post-infection, 90% of the animals had responded. This result is consistent with that obtained with the "acute" serum panel (Panel 2, Tufts), which showed 80% or higher of the patients whose serum had been collected 3 or more weeks after disease onset were positive by the C₆ ELISA. With the CDS serum panel of 40 specimens from convalescent patients, the sensitivity of the C₆ ELISA (85%) was slightly higher than that of the conventional ELISA (80%) and higher than the combination of IgG and IgM Western blot (75%). A lower sensitivity was obtained with serum specimens collected during the acute phase of Lyme disease (74%), with the 30 samples from Tufts-New England Medical Center. However, only 3 of these patients remained negative in the convalescent phase. The fourth negative sample among the convalescent phase samples had been positive when obtained during the acute phase. The possibility of making an earlier diagnosis by testing IgM response to C₆ both in monkeys and in humans was explored. In either case, no IgM antibody response was seen in the

absence of an IgG response, no matter how early after infection or disease onset the serum samples had been collected.

Overall, and as expected from the results obtained with rhesus monkeys as infection progresses in these animals, sensitivity of anti-C₆ antibody detection was higher in humans with later forms of Lyme disease. For patients in the early disseminated phase (neuroborreliosis), sensitivity was 95%, and 100% for patients with late Lyme disease (Table 3). The exception were the samples from patients with post-treatment Lyme disease syndrome, which were only 62% positive (Table 3). Patients in this group had a history of Lyme disease and despite having received antibiotic therapy, they had persistent Lyme disease symptoms. The etiology of these symptoms is currently under investigation at the NIH.

C. *Specificity of C₆ ELISA*

The C₆ ELISA yielded a specificity of 100% when serum samples from patients with other chronic infections or autoimmune diseases were tested (Table 4). This panel included 9 specimens from relapsing fever patients (CDC), 22 syphilis patient samples (10 from NIH and 12 from UCLA), and 46 specimens from patients with either multiple sclerosis, positive anticardiolipin antibody, positive rheumatoid factor, positive rapid plasma reagin, positive antinuclear antibody, Guillain-Barré syndrome or mycobacterial infection (NIH). Only two potential false positive results were obtained when a group of 99 human serum samples randomly collected at a local hospital in Louisiana, USA, were tested (Table 4). Lyme disease is not endemic in Louisiana, but the identity of the patients and their clinical history is unknown.

In the following table, these abbreviations are used: MS, multiple sclerosis; ACA, positive anticardiolipin antibody; RF, positive rheumatoid factor; RFR, positive rapid plasma reagin; ANA, positive antinuclear antibody; GBS, Guillain-Barré syndrome; Myco, mycobacterial infection.

Table 4

Specificity of C₆ ELISA

Sample Description	Sample No.	Positive	Negative	Specificity (%)
Relapsing fever	9	0	9	100
Syphilis (UCLA)	12	0	12	100
Syphilis (NIH)	10	0	10	100
MS	10	0	10	100
ACA	10	0	10	100
RF	10	0	10	100
ANA	10	0	10	100
GBS	1	0	1	100
Myco	5	0	5	100
Hospitalized patient	99	2	97	98
Overall	176	2	174	99

This data demonstrates that the C₆ ELISA is remarkably specific (Table 4). It could discriminate between Lyme borreliosis and infections with spirochetes of different species but the same genus (*B. hermsii*) or family (*T. pallidum*). Moreover, none of the samples from patients with autoimmune diseases or diseases that often need to be differentially diagnosed with respect to Lyme disease, such as multiple sclerosis or the Guillain-Barré syndrome were positive with the C₆ test. Only 2 of 99 serum samples from a local hospital in Louisiana, where Lyme disease is not endemic, yielded a positive C₆ ELISA result. Given that the samples were collected randomly from unknown patients, it is impossible to assess whether these two patients could have been exposed to *B. burgdorferi*. The specificity results obtained are supported by our finding that, except for the *Vls* cassette of Lyme spirochetes, no other sequences homologous to C₆ could be identified (using the

BLAST search algorithm) in the National Center for Biotechnology Information protein sequence data base. High diagnostic specificity is extremely critical to improve the positive predictive value of a test, especially when the incidence of a disease is very low. The prevalence of Lyme disease in most endemic areas is less than 0.01%;
5 the highest incidence rate in the US is 0.09% in Connecticut [Lightfoot et al., Ann Intern Med., 119:503-509 (1993); Tugwell et al., Ann Intern Med., 127:1109-1123 (1997)]. The overall specificity of the C₆ ELISA was 99% and the positive and negative predictive values were 99% and 89%, respectively.

The cassette portion of the VlsE contains 6 invariable regions
10 (IR) interspersed with an equal number of variable regions [Zhang et al., Cell, 89:275-285 (1997)]. The latter, by their very nature, have no diagnostic value. The additional 5 IRs are not as conserved as IR₆. Nonetheless, we assessed whether the remaining IRs (IR₁₋₅) could contribute to improve the diagnostic performance of C₆, whose sequence is based on that of IR₆. No antibody responses to peptides reproducing the
15 sequences of IR₁₋₅ were detected in humans or monkeys infected with *B. burgdorferi* (data not shown). On this basis we concluded that no improvement would be accrued from incorporating any of these peptides into the assay.

As expected, monkey serum samples which contained high-titer anti-OspA antibody did not react with C₆. This ELISA, therefore, is suitable in the
20 OspA-vaccine sera. Moreover, because of its simplicity and high specificity and sensitivity, the ELISA of the present invention alleviates the so-far intractable problems of Lyme disease serodiagnosis.

Example 9 - Diagnosis of Lyme Disease Using ELISA of Invention Regardless of Infecting Borrelia Genospecies

25 Serodiagnosis of Lyme disease is in part hampered by polymorphism of antigenic proteins of *B. burgdorferi*. In Europe this problem is more serious since all of the three pathogenic genospecies are prevalent. The studies provided in this examples section demonstrate that the VlsE immunodominant IR₆ is antigenically

conserved among the three *B. burgdorferi* genospecies and can serve as a universal probe for the serodiagnosis of Lyme disease. In this example, European strains were investigated.

A. *Human serum sources*

5 Two sources of human serum were utilized in this study, one from Austria and the other from Italy. Each source provided two panels, one from patients with early Lyme disease and the other from patients with late disease manifestations. The early serum panel from Austria was composed of 29 specimens serially collected from 11 patients with erythema migrans (EM). A commercial ELISA kit was used as
10 per the manufacturer's instructions (DAKOPATTS A/S, Copenhagen, Denmark) to test the serum specimens from Austrian patients with early Lyme disease. The antigen was native flagellar protein purified from cultured *B. afzelii* strain DK-1. A commercial immunoblot assay kit (MRL Diagnostics, Cypress, CA) was employed to analyze the Austrian serum panel obtained from patients with early Lyme disease,
15 according to the manufacturer's recommendations. The antigen consisted of whole-cell sonicates of cultured *B. garinii* strain 20047. A test was considered positive when at least one of the 23- or 39-kDa IgM bands or four of the 21-, 23-, 37-, 39-, 41-, 45-, or 93-kDa IgG bands were visible on the immunoblots. Austrian patients with early Lyme disease whose skin biopsies were cultured (all but one), were culture positive.
20 The late serum panel from this source consisted of 21 specimens from 21 IgG seropositive patients with clinically and histologically diagnosed acrodermatitis chronica atrophicans (ACA).

The Italian panel of early serum specimens consisted of 20 samples collected serially from 13 EM patients with culture confirmed *B. burgdorferi*
25 infection. To analyze serum specimens from the early Lyme disease Italian patients with increased sensitivity, three local strains *B. garinii* BITS, *B. afzelii* BL3 and *B. burgdorferi* sensu stricto Alcaide were individually utilized as a source of immunoblot antigens. The anti-*B. burgdorferi* antibodies were probed with goat anti-human IgG or IgM as secondary antibodies. A diagnosis was made based on criteria described
30 previously [U. Hauser et al, *J. Clin. Microbiol.*, 35:1433-1444 (1997); M. Cinco et al,

FEMS Immunol. Med. Microbiol., 12:217-222 (1995)]. Chronic serum specimens from Italy were from 20 patients with clinically diagnosed signs and symptoms that were consistent with late neuroborreliosis, arthritis or ACA. Early specimens were collected serially starting at disease onset and continuing thereafter for 3 months, during and after antibiotic treatment.

B. *Peptide-Based ELISA*

C₆ (CMKKDDQIAAAMVLRGMAKDGQFALK, SEQ ID NO:8), was synthesized and conjugated to biotin as described in Example 1E above. The ELISA was performed as described in Example 1G, using the human sera samples of Example 9A. The antigen-antibody reaction was probed using a peroxidase substrate system and optical density (OD) was measured at 450 nm.

C. *Sensitivity of the C₆ ELISA in patients with early Lyme disease*

Twenty of 23 patients with culture-confirmed EM (87%) had a detectable anti-IR₆ antibody response. One patient (A3) with positive EM was C₆ ELISA negative. No culture was performed with this patient. Results obtained with standard ELISA and immunoblots indicate that the C₆ ELISA is more sensitive than any of these tests, used either alone or combined, for detecting an early infection (Austrian panel). The C₆ ELISA also performed, in terms of diagnostic sensitivity, better than any of the three antigen preparations that were used to diagnose the Italian serum panel. Bacterial isolation and classification revealed that the patients were infected with either *B. garinii* or *B. afzelii* strains. This result provided additional evidence that IR₆ is antigenically conserved across the genospecies barriers of the *B. burgdorferi* sensu lato complex.

D. *Sensitivity of the C₆ ELISA for serodiagnosing late Lyme disease*

Twenty of 21 patients (95%) with histologically confirmed ACA and positive standard IgG serology showed a strong antibody response to IR₆. A sensitivity of 70% (14/20) was obtained when a panel of sera from patients with signs and/or symptoms consistent with late Lyme disease was tested. These 14 positive results included 3 (3/4) patients with late arthritis, 8 (8/12) with late neurologic manifestations, and 3 (3/4) with ACA, indicating that the C₆ ELISA was able to

detect different manifestations that might be caused by various *B. burgdorferi* strains. However, it should be pointed out that clinical diagnosis of late Lyme disease can be difficult due to the absence of well defined criteria.

E. Discussion

5 Experimental infections of monkeys and mice with *B. burgdorferi* sensu stricto strains B31, JD1, NT1 or Sh-2-82 have been found to elicit an early, strong and persistent antibody response to IR₆. All of these strains had been isolated in the United States. In another experiment, mice responded strongly to IR₆, regardless of whether they were infected with *B. garinii* or *B. afzelii* strains by tick or needle
10 inoculation. In the study described in this Example, sensitivities of 83% (20/24) and 95% (20/21) were obtained for European patients with clinically well defined early (EM, culture confirmed) and late (histologically confirmed) Lyme disease (ACA), respectively.

Several independent groups have noted that European patients show
15 restricted antibody responses to *B. burgdorferi* antigens, which in part contributes to the relatively lower sensitivity in European serodiagnosis. This difference was not apparent when the C₆ ELISA was used, as similar diagnostic sensitivities were obtained with North American [see earlier examples] and European serum samples. The exception was the panel of serum samples collected from Italian patients with
20 chronic symptoms, with which the C₆ ELISA yielded a sensitivity of only 70%. Clinical diagnosis of late Lyme disease is compounded by the absence of pathognomonic signs and the virtual impossibility of confirming diagnosis by culture.

Serodiagnosis of Lyme disease is further hampered by the low
specificity of the existing assays. This may be caused by crossreactive antigens shared
25 among *B. burgdorferi* and other pathogenic or nonpathogenic bacteria. Among these antigens are homologues of the omnipresent bacterial heat-shock proteins, flagellin, which is shared with other spirochetes such as *Borrelia hermsii* and *Treponema pallidum*, and a 60-kDa antigen which is expressed by a wide range of bacteria. Other bacterial infections may thus elicit antibodies that react with *B. burgdorferi*

antigens, causing false positive results. In contrast to conventional diagnostic techniques, the C₆ ELISA is highly specific. None of the seventy seven serum samples from patients infected with *B. hermsii*, the spirochete that causes relapsing fever, or *T. pallidum*, the agent of syphilis, or suffering from other chronic infections or diseases such as tuberculosis and multiple sclerosis, contained detectable antibody to C₆. Moreover, except for *B. burgdorferi* VlsE, no other protein sequences homologous to IR₆ could be identified by BLAST searches in the National Center for Biotechnology Information.

Recently, the initiation of Lyme disease immunoprophylaxis in humans through the use of the OspA (outer surface protein A) vaccine has made obsolete those diagnostic assays based on whole-cell antigens, as these preparations include OspA. Predictably, the C₆ ELISA does not detect anti-OspA antibodies. Hence, approval and use of the OspA vaccine in Europe will not affect the performance of the ELISA described herein. Further, the ELISA of the invention is readily adapted to yield a strictly quantitative readout, or to a more rapid type with a semi-quantitative readout.

Example 10 - Peptide-Based ELISA For Diagnosis of Canine Lyme Disease

Borrelia burgdorferi, the etiologic agent of Lyme disease, is able to infect a variety of mammalian species, including dogs. Of the domestic animals, dogs are at the greatest risk of becoming infected with *B. burgdorferi* and have been recommended as sentinel animals for human Lyme disease. Due to the lack of differential signs such as erythema migrans in infected dogs, laboratory methods are very important for canine Lyme disease diagnosis. In this study, sera collected from dogs experimentally infected with *B. burgdorferi* by tick inoculation were analyzed for an antibody response to the six invariable regions (IRs) of VlsE, the variable surface antigen of *B. burgdorferi*, by peptide-based enzyme-linked immunosorbent assays (ELISAs). Two IRs, IR₂ and IR₆, were found to be immunodominant. Serial immune response studies revealed that the antibody response to IR₆ appeared earlier and was stronger than the response to IR₂, suggesting that IR₆ alone was sufficient for

serodiagnosis. The IR₆ (C₆) peptide-based ELISA, when C₆ alone was used as antigen, showed a significant antibody response in 35% (7/20) dogs as early as 3 weeks post-infection. All dogs (n=33) became strongly positive one week later, and this response persisted for the entire study, which lasted 18 months. When both C₂ and C₆ combined were used as ELISA antigens the sensitivity for detecting specific antibody in both experimentally infected and/or clinically positive dogs could not be further improved. These results ruled out the necessity to include C₂ as diagnostic antigen. In addition, C₆ alone gave a similar sensitivity as the conventional immunoblotting or ELISA diagnosis when clinical samples were tested. Moreover, the C₆ ELISA resulted in 100% specificity with blood samples collected from 70 healthy dogs from an area where Lyme disease is not endemic, 13 dogs with other infections, and 10 animals vaccinated with OspA or whole-spirochete vaccines. Therefore, this single probe can be sensitive and specific for canine Lyme disease serodiagnosis.

All above-noted references and priority document are incorporated herein by reference. Numerous modifications and variations of the present invention are included in the above-identified specification and are expected to be obvious to one of skill in the art. Such modifications and alterations to the compositions and processes of the present invention are believed to be encompassed in the scope of the claims appended hereto.

WHAT IS CLAIMED IS:

1. A peptide IR₆, MKKDDQIAAAMVLRGMAKDGQFALKD
SEQ ID NO: 1, isolated from cellular materials with which it is naturally associated.
2. An artificial protein selected from the group consisting of:
 - (a) a protein comprising the amino acid sequence selected from the group consisting of MKKDDQIAAAMVLRGMAKDGQFALKD SEQ ID NO: 1 (IR₆) or an analog, homolog or fragment thereof; and
 - (b) a fusion protein comprising the amino acid sequence of (a) and a fusion partner; and
 - (c) a deletion protein comprising the amino acid sequence of (a) with one or more amino acids deleted therefrom.
3. The protein according to claim 2, wherein the fragment of IR₆ is a T or B cell epitope.
4. The protein according to claim 2, wherein the fusion protein comprises a T or B cell epitope of IR₆ fused to a selected fusion partner.
5. The protein according to claim 2, wherein the fusion partner is selected from the group consisting of:
 - (a) IR₁, GNAAIGDVV SEQ ID NO: 3
 - (b) IR₂, SVNGIAKGIKIVDAA SEQ ID NO: 4
 - (c) IR₃, AGKLFVK SEQ ID NO: 5,
 - (d) IR₄, DAGKAAAAVAASGEQILKAIVHAA SEQ ID NO: 6,
 - (e) IR₅, ATNPIDAAIG SEQ ID NO: 7; and
 - (f) and analogs, homologs, and derivatives of IR₁-IR₅.

6. A nucleic acid sequence encoding IR₆,
MKKDDQIAAAMVLRGMAKDGQFALKD SEQ ID NO: 1 or a fragment thereof.
7. A nucleic acid sequence selected from the group consisting of:
 - (a) a nucleic acid sequence which hybridizes to the sequence of claim 6 under stringent conditions;
 - (b) a nucleic acid sequence complementary to the sequence of claim 6;
 - (c) a nucleic acid sequence encoding a fusion molecule comprising MKKDDQIAAAMVLRGMAKDGQFALKD SEQ ID NO: 1 or a fragment thereof and a fusion partner; and
 - (d) an allelic variant of any of (a) through (c).
8. A vector comprising a nucleic acid sequence according to claim 6 or 7 under the control of suitable regulatory sequences.
9. A host cell transformed with the vector according to claim 8.
10. A diagnostic reagent comprising a nucleic acid sequence of claim 6 or 7 and a detectable label which is associated with said sequence.
11. A method for specifically diagnosing Lyme disease, said method comprising the steps of performing PCR using a nucleic acid sequence according to claim 6 or 7 or diagnostic reagent according to claim 10.
12. A diagnostic reagent comprising a peptide according to claim 1 or a protein according to claim 2 and a detectable label which is associated with said peptide or protein.

13. A method for specifically diagnosing Lyme borreliosis in a human or animal comprising the steps of:

contacting a sample from a human or animal to be diagnosed with the peptide of claim 1, the protein of 2, or the diagnostic reagent of claim 12, whereby in the presence of antibodies to *B. burgdorferi* in the sample, an antibody complex is formed with the peptide, protein or reagent, and

analyzing for the presence of said complex.

14. The method according to claim 13, wherein the method is an enzyme linked immunosorbent assay (ELISA).

15. An isolated antibody which is specific for the peptide of claim 1 or a fragment thereof.

16. The antibody according to claim 15, isolated by immunizing a host with the peptide of claim 1 or the protein of claim 2.

17. The antibody according to claim 15 which is selected from the group consisting of a chimeric antibody, a humanized antibody, a monoclonal antibody and a polyclonal antibody.

18. A diagnostic reagent comprising the antibody according to claim 15 and a detectable label.

19. A method of specifically diagnosing Lyme disease in a human or an animal, said method comprising the steps of:

contacting the antibody of claim 15 or diagnostic reagent of claim 18 with a sample from a human or animal to be diagnosed, whereby in the

presence of *B. burgdorferi*, a complex is formed with the antibody or diagnostic reagent, and

analyzing for the presence of said complex.

20. An anti-idiotypic antibody specific for the antibody of claim 15.

21. A diagnostic reagent comprising an anti-idiotypic antibody according to claim 20 and a detectable label linked thereto.

22. A method of specifically diagnosing lyme disease in a human or an animal, said method comprising:

contacting the antibody of claim 20 or diagnostic reagent of claim 21 with a sample from a human or animal to be diagnosed, whereby in the presence of *B. burgdorferi*, a complex is formed with the antibody or diagnostic reagent, and

analyzing for the presence of said complex.

23. A kit for diagnosing infection with *B. burgdorferi* in a human or animal comprising a diagnostic reagent according to any of claims 12, 18 or 21.

24. A composition comprising an effective amount of a peptide according to claim 1 or a protein according to claim 2 and a pharmaceutically acceptable carrier.

25. The composition according to claim 24, wherein the composition contains a fusion protein comprising of a fragment of MKKDDQIAAAMVLRGMAKDGQFALKD SEQ ID NO: 1 consisting of a T cell epitope fused to a fusion partner.

26. The composition according to claim 25, wherein the fusion partner is a peptide selected from the group consisting of:

- (a) IR₁, GNAAIGDVV SEQ ID NO: 3
- (b) IR₂, SVNIGAKGIKGIVDAA SEQ ID NO: 4
- (c) IR₃, AGKLFVK SEQ ID NO: 5,
- (d) IR₄, DAGKAAAVA AVS GEQILKAIVHAA SEQ ID NO: 6,
- (e) IR₅, ATNPIDAAIG SEQ ID NO: 7; and
- (f) and analogs, homologs, and derivatives of IR₁-IR₅.

27. The composition according to claim 24 wherein said composition comprises at least one other *B. burgdorferi* antigen or fragment thereof.

28. The composition according to claim 27 wherein said other antigen is selected from the group consisting of OspA, OspB, OspC, BmpA, BmpB, BmpC, BmpD and fragments or variants thereof.

29. A method of vaccinating a human or animal against Lyme Disease comprising administering to said human or animal a composition comprising an effective amount of the composition of claim 24.

30. A method for treating Lyme Disease in a vertebrate host comprising administering an effective amount of a composition according to claim 24.

31. A method of identifying compounds which specifically bind to an epitope in IR₆, MKKDDQIAAAMVLRGMAKDGQFALKD SEQ ID NO: 1, comprising the steps of contacting said IR₆ with a test compound to permit binding of the test compound to IR₆; and determining the amount of test compound which is bound to IR₆.

32. A compound identified by the method of claim 31.
33. A vaccine composition comprising:
- (a) a peptide selected from the group consisting of:
- (i) IR₁, GNAAIGDVV SEQ ID NO: 3
 - (ii) IR₂, SVNGIAKGIGIVDAA SEQ ID NO: 4
 - (iii) IR₃, AGKLFVK SEQ ID NO: 5,
 - (iv) IR₄, DAGKAAA VAAVSGEQILKAIVHAA
SEQ ID NO: 6,
 - (v) IR₅, ATNPIDAAIG SEQ ID NO: 7; and
 - (vi) and analogs, homologs, and derivatives of IR₁-
IR₅; and
- (b) a pharmaceutically acceptable carrier.
34. A vaccine composition comprising:
- (a) a nucleotide sequence encoding a peptide selected from the
group consisting of:
- (i) IR₁, GNAAIGDVV SEQ ID NO: 3
 - (ii) IR₂, SVNGIAKGIGIVDAA SEQ ID NO: 4
 - (iii) IR₃, AGKLFVK SEQ ID NO: 5,
 - (iv) IR₄, DAGKAAA VAAVSGEQILKAIVHAA
SEQ ID NO: 6,
 - (v) IR₅, ATNPIDAAIG SEQ ID NO: 7; and
 - (vi) and analogs, homologs, and derivatives of IR₁-
IR₅; and
- (b) a pharmaceutically acceptable carrier.

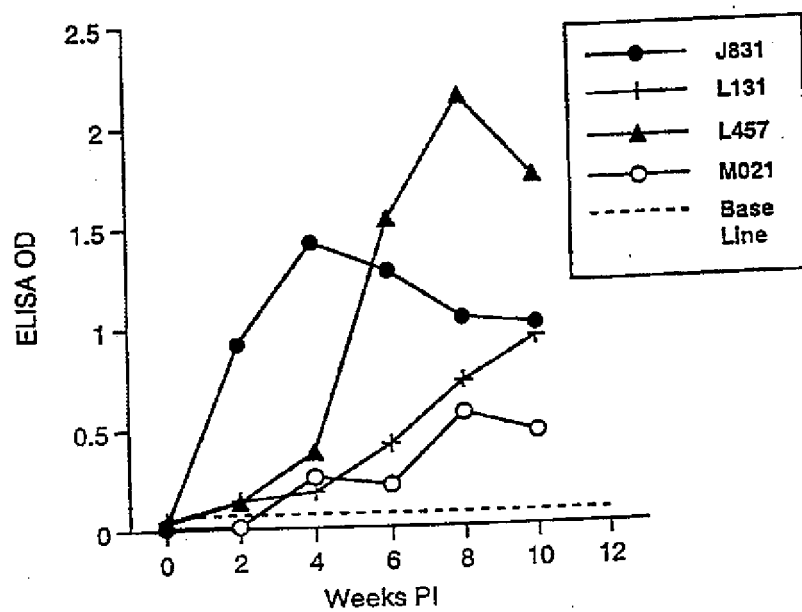


Fig. 1

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FIGURE 2

	10	20	30	40	50	60
P7-1	KNNDHKNHKG	TVKNAVDMAK	AAEEAASAAS	AATGNAAIGD	VVKNSGAAAK	GGEAASVNGI
B31 Vls	A*S GEQILSA IVKA**AG*A DQD*EKPG-* AKNP IA**IG K*N*DDGADF					

	70	80	90	100	110	120
P7-1	ARGIKGIVDA	AGKADAKEGK	LDATGAEGTT	NVNAGKLFVK	RAADDGGDAD	DAGKAAAAVA
297 Vls						ATE**TA*
B31 Vls	GD*M*K-D*Q	IAA*I*LR*M	-AKD*KFAVK	KDEK**A--E	<u>G*IKGASELL</u>	*KLVK*VKT*

	130	IR ₁	140	150	IR ₂	160	170	180
P7-1	ASAATGNAAI	GDVVNGDVAK	AKGGDAABVN	GIKGIKGI	DAIEKADAKE	GKLNAAGAEG		
297 Vls	SGDKEMIGKV	VK*TNACA*A	****EEK**	****S*****	E*****G--*	***E**AGD*		
B31 Vls	EG*SS*T**	*E** <u>DN-A-A</u>	-*AA* <u>KD**T</u>	*****E**	<u>E**G--GSE-</u>	<u>K-KV*A*K*</u>		
			VR _I			VR _{II}		

	190	IR ₃	200	210	IR ₄	220	230	240
P7-1	TTNADAGKLE	VKNAGNVGGE	AGDAGKAAVA	VAAVS GEQIL	KALVHAAKDG	GEKQGKKAAD		
297 Vls	NKD*C*****	A***A*G**G	*AA*E*****	*S***K***	*****D**G-K	E**GVADVKE		
B31 Vls	<u>EN*KG*****</u>	<u>G*AGA*AH*D</u>	SEA*S***G*	*S*****	<u>S***K**GEA</u>	<u>*DQE***PEE</u>		
		VR _{III}				VR _{IV}		

	IR ₅	250	260	270	IR ₆	280	290	300
P7-1	ANNPDAATG	GAGDNDAAAA	FATMKKDDOI	AAAMVLRGMA	KDGOFALKDA	AAAHEGTVKN		
297 Vls	*****E****	ST*****F	QDE***N***	***I*****	***E*****N	EHDKAKGL*S		
B31 Vls	<u>*K***A****</u>	<u>DKD-G**EFN</u>	<u>QDG*****</u>	<u>***IA*****</u>	<u>***K**V**G</u>	<u>GEKA**AI*G</u>		
		VR _V				VR _{VI}		

	310	320	330	340
P7-1	AVDIKAAAE	AASAASAATG	SAAIGDVVNG	NGATAKGGDA KSVNGIAKG
297 Vls	T*E			
B31 Vls	VSELLDLVKA	VKT*EK*SS*	T****E**A-	DA-A-*VA*K A**T*****

Fig. 3A

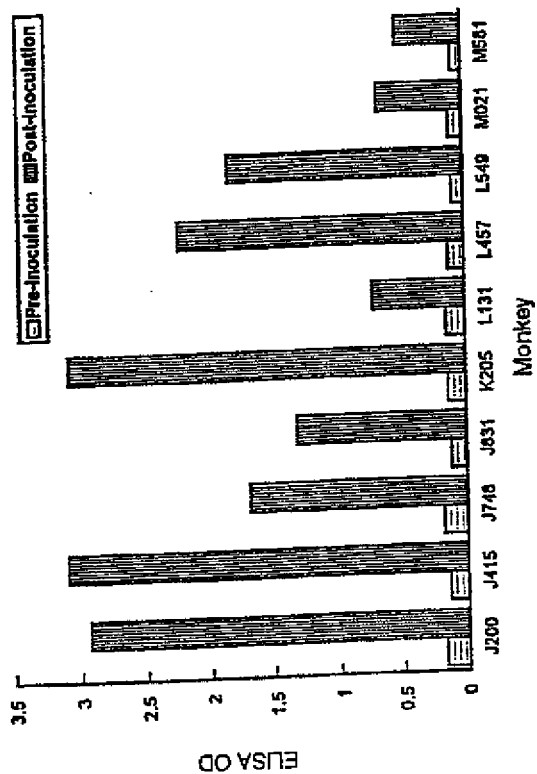


Fig. 3B

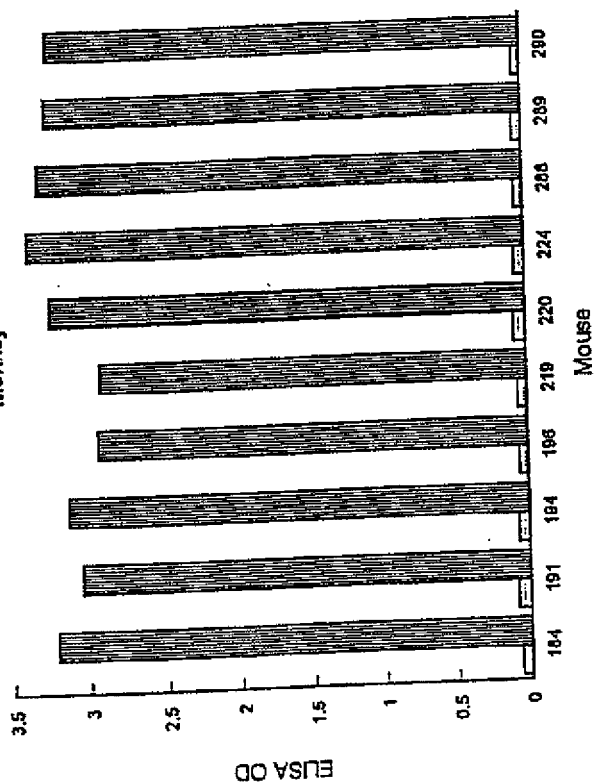


Fig. 4A

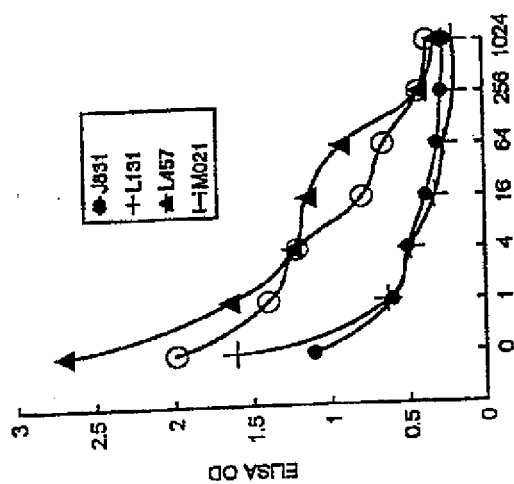
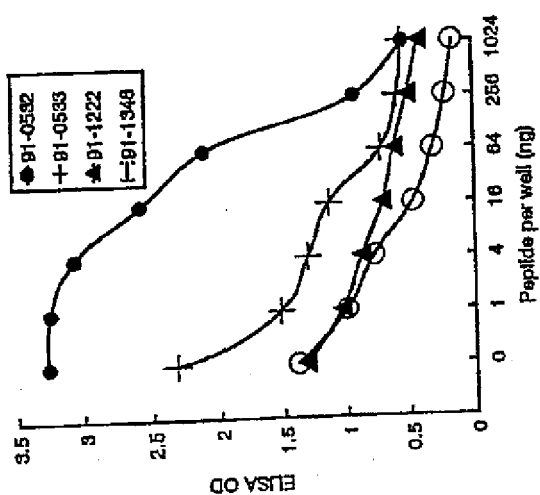


Fig. 4B



SEQUENCE LISTING

<110> The Administrators of the Tulane Educational Fund
Philipp, Mario T.
Liang, Fang Ting

<120> Peptides and Assays for the Diagnosis of Lyme Disease
and Compositions Useful for the Prevention Thereof

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20 25

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<400> 8

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1 5 10 15

Met Ala Lys Asp Gly Gln Phe Ala Leu Lys
 20 25

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 00/11085

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12N15/31 C07K14/00 C07K14/20 C07K16/12 G01N33/68
A61K39/02 A61K38/00 C12N15/62

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 C07K C12N A61K G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, CHEM ABS Data, WPI Data, CAB Data, PAJ, BIOSIS, MEDLINE, STRAND

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 99 00413 A (UNIV TULANE ;PHILIPP MARIO T (US)) 7 January 1999 (1999-01-07) the whole document	1-34
P,X	F.T. LIANG ET AL.: "Analysis of antibody response to invariable regions of VlsE, the variable surface antigen of <i>Borrelia burgdorferi</i> . " INFECTION & IMMUNITY, vol. 67, no. 12, December 1999 (1999-12), pages 6702-6706, XP000925195 the whole document	1-34

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

7 September 2000

Date of mailing of the international search report

14/09/2000

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Hix, R

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 00/11085

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>HANSEN K ET AL: "IMMUNOCHEMICAL CHARACTERIZATION OF AND ISOLATION OF THE GENE FOR A BORRELIA BURGDORFERI IMMUNODOMINANT 60-KILODALTON ANTIGEN COMMON TO A WIDE RANGE OF BATERIA" INFECTION AND IMMUNITY,US,AMERICAN SOCIETY FOR MICROBIOLOGY. WASHINGTON, vol. 56, no. 8, 1 August 1988 (1988-08-01), pages 2047-2053, XP002001271 ISSN: 0019-9567 cited in the application the whole document</p> <p>---</p>	
A	<p>SIMPSON W J ET AL: "REACTIVITY OF HUMAN LYME BORRELIOSIS SERA WITH A 39-KILODALTON ANTIGEN SPECIFIC TO BORRELIA BURGDORFERI" JOURNAL OF CLINICAL MICROBIOLOGY, WASHINGTON, DC, US, vol. 28, no. 6, June 1990 (1990-06), pages 1329-1337, XP000925133 ISSN: 0095-1137 cited in the application the whole document</p> <p>---</p>	
A	<p>BARBOUR A G ET AL: "VARIATION IN A MAJOR SURFACE PROTEIN OF LYME DISEASE SPIROCHETES" INFECTION AND IMMUNITY, AMERICAN SOCIETY FOR MICROBIOLOGY. WASHINGTON, US, vol. 45, no. 1, July 1984 (1984-07), pages 94-100, XP000915468 ISSN: 0019-9567 cited in the application the whole document</p> <p>---</p>	
A	<p>EP 0 465 204 A (AMERICAN HOME PROD) 8 January 1992 (1992-01-08) cited in the application the whole document</p> <p>---</p>	
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Information on patent family members

International Application No

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